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Review

The Application of Epicardium in Heart Failure Treatment: Opportunities and Challenges

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Abstract

Heart failure remains_one of the leading causes of morbidity and mortality worldwide. Conventional treatment strategies, while beneficial, face numerous limitations. Drug therapies may lead to resistance, while device-based treatments such as LVAD and ICD carry risks of infection, bleeding, device failure, and high costs. For end-stage heart failure, heart transplantation is further constrained by donor shortages and immune rejection. In contrast, cell-based therapies have emerged as a promising alternative. Recent studies have highlighted the critical role of the epicardium and epicardium-derived cells (EPDCs) in cardiac regeneration. These cells contribute to heart repair through multiple mechanisms, including direct cell therapy, the development of epicardium-based biomaterials, and integration with gene therapy approaches. This review outlines the anatomical structure and biological functions of the epicardium, explores the regenerative potential of the epicardium and EPDCs, and evaluates their application in heart failure treatment. Furthermore, it discusses the translational potential and current challenges associated with epicardial-based therapies, offering novel insights and strategies for heart failure management.

Introduction

Heart failure (HF) is a common heterogeneous clinical syndrome characterized by high morbidity and mortality. It significantly impairs patients' quality of life and contributes to a substantial global health burden. Although the incidence of HF has stabilized or declined in high-income countries in recent years, the global prevalence continues to rise due to population aging and the increasing prevalence of risk factors, with a concerning trend toward younger onset[1]. The pathogenesis of HF involves a variety of factors, including ventricular systolic or diastolic dysfunction, abnormalities, and disturbances in cardiac rhythm or conduction[2]. Current therapeutic strategies for HF include pharmacological interventions, lifestyle modifications, surgical procedures, and heart transplantation. However, these approaches demonstrate limited efficacy in heart failure with preserved ejection fraction (HFpEF) and are further

challenged by issues such as drug resistance, adverse effects, and high treatment costs[3].

Cardiac regeneration relies on the generation of functional cardiomyocytes and the formation of a vascular network to support these newly formed cells[4]. In recent years, the epicardium has garnered increasing attention as a critical tissue in cardiac regeneration. Rich in progenitor cells, the epicardium plays a pivotal role in signal transduction during heart repair[5]. A subset of EPDCs has been shown to differentiate into cardiac fibroblasts and vascular smooth muscle cells, thereby providing essential structural and functional support myocardium[6]. Consequently, the epicardium holds significant therapeutic promise for HF.

Nevertheless, despite growing interest in the role of the epicardium in HF treatment, several challenges remain. Key among these are the need to effectively activate epicardial cells to promote myocardial regeneration[6], and to address the potential for fibrosis during epicardial repair processes[7, 8]. This review aims to explore the therapeutic potential of the epicardium in HF, analyze its role in myocardial repair, and discuss the challenges and future directions of epicardial-based therapies.

Biological Functions and Regenerative Potential of the Epicardium

The pericardium is anatomically divided into the fibrous pericardium and the serous pericardium. The latter consists of a parietal layer and a visceral layer, with the visceral layer constituting the epicardium[9]. The epicardium is composed primarily of a single layer of flattened epithelial cells, which connect to the underlying myocardium through layer a connective tissue[10]. During embryonic development, the epicardium serves two essential roles: it acts as a source of progenitor cells for various cardiac lineages, and it provides a paracrine environment conducive to cardiomyocyte compaction and maturation[11]. During development, a subset of epicardial cells undergoes epithelial-to-mesenchymal transition (EMT) and migrates into the myocardium, differentiating into various cell types that are essential for normal heart development (Figure 1). In addition, epicardial cells contribute to cardiac repair by generating multiple cell types and secreting paracrine factors.

Notably, the epicardium plays a central role in myocardial repair through multiple mechanisms. Its close interaction with the myocardium enables epicardial-derived signals to directly stimulate cardiomyocyte proliferation, while signals from cardiomyocytes induce EMT in epicardial cells, facilitating their differentiation into cardiomyocytes and coronary vascular cells[13]. Lavine et al. demonstrated that fibroblast growth factor (FGF) signaling between the epicardium and myocardium activates Hedgehog (Hh) signaling and induces endothelial vascular growth factor expression, thereby promoting coronary vessel formation[14, 15]. Huang et al. reported that suppression of CCAAT/enhancer-binding protein

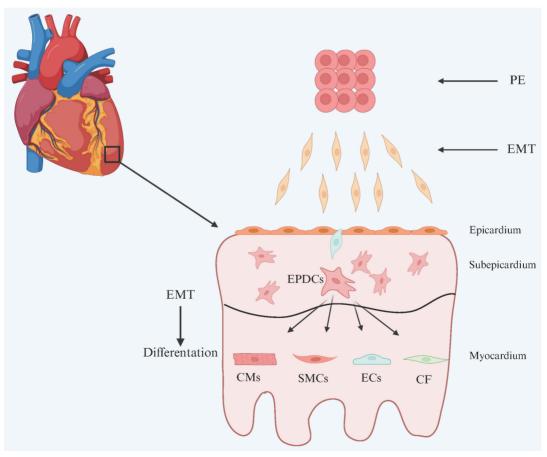


Figure 1. Development of the epicardium and its lineages. The epicardium originates from the proepicardium (PE). Proepicardial cells undergo epithelial-to-mesenchymal transition (EMT) and migrate to the myocardial surface of the looping heart, where they adhere, migrate, proliferate, and undergo mesenchymal-to-epithelial transition (MET) to form a squamous epithelial layer, the epicardium. While the epicardium remains an intact layer, some epithelial cells undergo a second round of EMT and migrate into the mustive-rich sub-epicardial layer. Fate mapping and genetic lineage tracing studies have demonstrated the multi-lineage potential of these EPDCs differentiate into smooth muscle cells (SMCs), contributing to the formation of coronary vasculature, and into cardiac fibroblasts (CF) in the mature heart. Although the contribution of EPDCs to cardiac endothelial cells (ECs) and cardiomyocytes (CMs) has been described, this remains a subject of ongoing debate. (The image has been adapted based on reference[12].

(C/EBP) signaling in the epicardium reduced fibrosis and improved contractile function post-myocardial infarction, likely by inhibiting neutrophil infiltration into the infarcted region[16]. Recent studies by Elizabeth et al. revealed that following ventricular amputation, neutrophils rapidly accumulate and growth factor ligand secrete the epidermal heparin-binding EGF-like growth factor (HBEGF), which acts in concert with other FGFs and MAPK/ERK pathways to induce epicardial regeneration and contribute to cardiac repair[17]. Additionally, Bargehr et al. found that exosomes derived from mouse epicardial cells contain microRNAs such as miR-30a, miR-100, and miR-27a, which promote the proliferation of human pluripotent stem cell-derived cardiomyocytes[18].

The epicardium harbors multipotent progenitor cell population that, upon undergoing EMT, loses cell-cell adhesion and apical-basal polarity while acquiring mesenchymal characteristics, migration and enabling invasion into myocardium. These cells are known as EPDCs. EPDCs can differentiate into coronary vascular smooth muscle cells, cardiac fibroblasts, endothelial cells, and other cell types[19-21]. During EMT, EPDCs acquire mesenchymal properties and express mesenchymal proteins. Eroglu et al. demonstrated in a salamander cryoinjury model that CLDN+ epicardial cells formed honeycomb-like structures via tight junctions at the injury site, underwent transcriptional reprogramming, and ultimately differentiated into cardiomyocytes[22].

However, in the adult heart, the epicardium remains quiescent and is only reactivated following cardiac injury. Compared to fetal development, the contribution of EPDCs to myocardial repair in adults is significantly reduced. Multiple studies have shown that most newly formed cells after injury originate from pre-existing cardiomyocyte subpopulations, rather than from newly differentiated EPDCs. Therefore, the role of epicardial cells in adult hearts is largely paracrine rather than serving as a direct source of reparative cells[16, 23-25]. EPDCs mostly remain on the heart surface and preferentially differentiate into cardiac fibroblasts and smooth muscle cells. They contribute to the survival and growth of coronary vessels after myocardial infarction through paracrine secretion of factors such as VEGFA and FGF2[26-28]. Moreover, unlike mesenchymal stem cells (MSCs), EPDCs uniquely express cardiac-specific marker genes such as GATA4 and cTnT. Comparative studies between primary human MSCs and EPDCs derived from the human right atrial appendage confirmed that only EPDCs exhibit these myocardial markers. Although EPDCs display osteogenic differentiation

potential *in vitro* under conditions that promote MSCs differentiation into adipogenic and osteogenic lineages, they fail to differentiate into adipocytes or endothelial cells[29].

Although EPDCs can reach injured sites and contribute to regeneration, their reparative capacity is often suppressed. As a result, EPDCs primarily exert their effects through the secretion of growth factors, cytokines, and extracellular matrix components that promote the proliferation, migration, differentiation of resident cardiac cells. EPDCs proliferation, migration stimulate the differentiation of resident cells by secreting growth factors, cytokines and extracellular matrix, thereby promoting tissue repair and regeneration. However, their regenerative potential is limited. Therefore, the epicardium is a powerful tool for new therapeutic strategies, which can synergistically repair damaged myocardium through differentiation into functional myocardium and endogenous response to paracrine stimulation[11]. EPDCs are known to secrete key factors such as retinoic acid and fibroblast growth factors, which are critical for cardiomyocyte proliferation. Moreover, GATA4 and GATA6 signaling within the epicardium regulates the development of the cardiac neural crest[30-32]. In zebrafish models, following acute cardiac injury, FGF signaling pathways are activated, with increased expression of FGFR2 and FGFR4 observed in EPDCs surrounding the atrium. FGFR4 remains highly expressed near the wound site even 30 days post-amputation. Inhibition of FGFR signaling in the epicardium disrupts the EMT process neovascularization in the injured region[33].

It is also noteworthy that the epicardium plays a crucial role in transport. During embryonic development, epicardial mesothelial cells actively transport fluids, cells, and particles across the serosa and pericardial cavity. Additionally, they synthesize and secrete mediators that regulate inflammation, immunity, and tissue repair[34].

Applications of the Epicardium in Heart Failure Therapy

The epicardium possesses a range of critical biological functions and remarkable regenerative potential, making it a key focus in recent heart failure research. As our understanding of epicardial development and regenerative properties deepens, researchers are exploring their therapeutic applications in cardiac repair from multiple perspectives. Currently, the therapeutic potential of the epicardium is mainly reflected in three areas: (1) the use of epicardial and EPDCs for cell therapy, leveraging their multipotent differentiation capacity

and paracrine effects to promote myocardial and vascular regeneration; (2) the development of epicardium-based biomaterials and tissue engineering scaffolds, providing mechanical support and a conducive microenvironment for cardiac repair; and (3) the use of the epicardium as a targeted delivery platform for gene therapy, aiming to enhance cardiac repair and functional recovery through the activation or editing of key signaling pathways (Figure 2). These strategies offer new avenues for precision therapy and clinical translation in the treatment of heart failure.

Epicardium-Based Cell Therapy

EPDCs play a pivotal role in cardiac repair. Studies have demonstrated that EPDCs exhibit diverse differentiation potentials under different stimuli (Table 1). For instance, atrial natriuretic peptide can induce EPDCs differentiation into adipocytes, whereas angiotensin II promotes their differentiation into myofibroblasts[35]. Overexpression of Cdx1 enhances EMT in epicardial cells, facilitating their differentiation into coronary vascular smooth muscle cells (SMCs) and endothelial cells, while also promoting their migration into the myocardium[36]. Thymosin β 4 (T β 4) has been shown to enhance the migratory capacity of adult EPDCs, thereby augmenting their regenerative potential[37]. In vitro studies using chick and mouse epicardial explants have revealed that TGF-β receptor stimulation can induce EMT and differentiation into SMCs. Similarly, stimulation with platelet-derived

growth factor (PDGF) receptors or FGF1/2/7 also promotes EMT and SMCs differentiation[5]. Dronkers *et al.* reported that EPDCs spontaneously undergo EMT *in vitro*; in adult EPDCs, this can be inhibited by the ALK5 kinase inhibitor SB431542, though its effect on fetal EPDCs is limited[38]. Furthermore, using an inducible large T-antigen (LT) expression system regulated by doxycycline, highly proliferative adult EPDCs can be obtained. Upon withdrawal of doxycycline, these cells retain epicardial morphology, EMT capacity, and paracrine signaling properties[39].

In adult mammals, the heart has limited capacity to regenerate functional cardiomyocytes after injury, often resulting in inflammation and fibrotic scar formation. The epicardium and its derivatives are central to this fibrotic process. Typically quiescent in the adult heart, epicardial cells can be reactivated following myocardial injury by re-expressing progenitor markers such as Raldh2, Tbx18, and WT1. These activated cells undergo EMT and generate EPDCs, which subsequently differentiate into multiple cell types, including SMCs, endothelial cells (promoting neovascularization), and fibroblasts 40-42]. (facilitating collagen deposition)[36, Additionally, the epicardium contributes to cardiac repair by recruiting regulatory T cells to suppress excessive immune responses, limiting pathological remodeling, and by secreting paracrine factors that stimulate coronary vessel growth and improve myocardial regeneration[42].

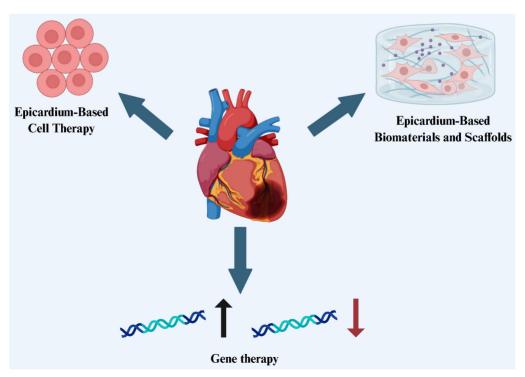


Figure 2. Therapeutic applications of the epicardium in heart failure

Table 1. Effects of Different Influencing Factors on EPDCs

Effects	Influencing factors	References
Differentiation potential under different conditions	 Atrial natriuretic peptide can promote its differentiation into adipocytes Angiotensin II promotes differentiation into myofibroblasts 	[5, 35, 36]
	 Overexpression of Cdx1 promotes differentiation into coronary artery SMCs and endothelial cells 	
	For TGF-β receptor promotes differentiation into SMCs	
	FGF1/2/7 can induce SMCs differentiation	
Influence EMT in epicardial cells	 Overexpression of Cdx1 c TGF-β receptor stimulation PDGF receptors and FGF1/2/7 ALK5 kinase inhibitor SB431542 inhibits EMT in adult EPDCs, but has little effect on fetuses 	[5, 36, 38]
Influence proliferation and migration	 Tβ4 has been shown to enhance the migratory capacity of adult EPDCs Inducible LT expression system regulated by doxycycline enables high proliferation capacity 	

Abbreviation: SMCs: smooth muscle cells; TGF-β: transforming factor β; PDGF: platelet-derived growth factor; FGF: fibroblast growth factor; LT: large T-antigen.

Nevertheless, the intrinsic response of EPDCs alone is insufficient for complete myocardial repair. For example, silencing intraflagellar transport protein 88 (IFT88), which is essential for ciliary function, promotes epicardial **EMT** and enhances Wnt/β-catenin signaling and bFGF expression. This leads to an increased number of WT1⁺ cells in both the epicardium and myocardium, facilitates EMT and angiogenesis after myocardial infarction, attenuates hypertrophic remodeling, and improves cardiac function[41]. Tcf21 is a critical target for regulating cardiac fibrosis. Loss of Tcf21 function in EPDCs enhances myofibroblast differentiation, while its overexpression suppresses this process. nanoparticles carrying IncRNA-TARID upregulate Tcf21 expression in EPDCs, significantly improving cardiac function and histological outcomes in both mouse and porcine myocardial infarction models[40]. Oxytocin activates its receptor via the TGF-β signaling pathway and its downstream effectors, reprogramming epicardial cells toward a progenitor-like state[43]. Moreover, Sca1+ bone marrow cells from young mice can promote EPDCs proliferation, migration, and EMT reactivation via the TGF-β pathway, thereby enhancing regeneration[44].

Epicardium is also a direct gateway for material exchange between pericardial fluid and myocardial tissue. The cells and molecular substances of pericardial fluid and myocardium can be freely interchanged through epicardium. The stem cells injected into the pericardial cavity can directly penetrate the epicardial mesothelial cells into the

myocardial tissue and play a therapeutic role in heart failure[45]. The delivery of extracellular matrix, growth factors, drugs, bioactive materials and genetic materials through the pericardial cavity can smoothly reach the myocardium through the epicardium and play a role in the treatment of myocardial diseases. Transcardial mesothelial cell route not only overcomes the limitations of traditional systemic drug delivery methods, but also has a variety of intrinsic advantages compared with other methods, which can achieve greater therapeutic effect. Intrapericardial drug delivery shows versatility in dealing with some cardiac diseases. Continuous research in this field undoubtedly brings further innovation and progress hope for improving cardiac therapy[46].

Collectively, these findings highlight the therapeutic potential of EPDCs in heart failure. By harnessing their multipotent differentiation capacity, EPDCs can promote myocardial regeneration, angiogenesis, and cardiac function recovery. Particularly, through the regulation of cell migration, EMT, and immune responses, EPDCs not only contribute to myocardial tissue repair but also enhance coronary vessel growth and mitigate pathological remodeling via paracrine signaling. These insights provide a promising foundation for the development of epicardium-based cell therapies in clinical heart failure treatment.

Epicardium-Based Biomaterials and Scaffolds

Spindle-shaped epicardium-derived cells secrete various extracellular matrix (ECM) components, such as osteopontin and fibronectin[47, 48]. When combined with biomaterials, these cells can be used as injectable materials to enhance cell adhesion and reduce apoptosis. The epicardium also provides essential **ECM** components for vascular reconstruction muscle regeneration[22]. and Epicardial composite patches fabricated from human cardiac extracellular matrix (hcECM) can be processed into hydrogels (hgECM) and coated onto scaffolds. These constructs maintain nanoscale structural stability and mechanical properties while improving cell adhesion and reducing cell necrosis on the scaffold surface[49]. When mechanically integrated with the heart as structural components of the cardiac chamber, epicardial-mimicking patches exhibit elastic properties similar to those of the native epicardium. After eight weeks of implantation in a rat myocardial infarction model, these patches demonstrated electrical signal conduction, reduced wall stress, and improved contractile function[50].

Reproducible activation of epicardial behavior has become a key therapeutic target in treating cardiac injury. Transitioning epicardial cells from an epithelial to a motile mesenchymal phenotype is crucial for their beneficial effects on the myocardium. The goal of engineered cardiac patches is to replicate the native ECM, provide mechanical support, and create a favorable environment for cell migration, integration, proliferation, and differentiation, while also facilitating efficient nutrient and waste exchange[51]. By suturing cardiac patches onto the epicardial surface, treatment areas can be precisely localized, enhancing therapeutic efficiency. When conditioned medium from epicardial cells is embedded into collagen nanofiber membranes and implanted into the heart, improvements myocardial contractility and prevention of excessive fibrosis after infarction have been observed. Wei et al. found that the expression of follistatin-like 1 (Fstl1) in the epicardium decreases following myocardial infarction. Application of a human Fstl1 epicardial patch stimulated cell cycle re-entry and proliferation of existing cardiomyocytes, thereby improving cardiac function and survival in both mouse and pig models[52].

Three-dimensional (3D) printing technology enables the precise construction of cardiac patches by depositing successive layers of biomaterials into a defined 3D structure. This technique allows for fine control over patch geometry, diameter, and pore size, promoting nutrient transport, enhancing proliferation, and ensuring adequate oxygen and nutrient supply within the scaffold[53, 54]. When designing cardiac patches, special attention must be paid to the anisotropic nature of the myocardium. In the longitudinal axis of the myocardium, intercalated discs and gap junction chains facilitate ionic current flow, while the transverse axis has fewer gap junctions, resulting in higher resistance. This significantly anisotropy affects myocardial conductivity. Improperly designed patches may exacerbate anisotropy and trigger arrhythmias[55].

Given the contractile-relaxation cycle of cardiac activity, the elasticity of the patch is equally critical. Patch materials must exhibit excellent biocompatibility to support cell adhesion and growth, low immunogenicity, and a degradation rate matched to the pace of tissue remodeling. The patch's electrical conductivity should approximate that of native myocardium to ensure synchronized contraction and electrical pacing. Cardiac patches not only provide mechanical support to infarcted areas but also serve as a delivery platform for cells or bioactive factors, thereby improving cell retention and enhancing therapeutic efficacy. Although cell transplantation using patches has shown promise in myocardial repair, the number of surviving cells and their integration with infarcted tissue remain suboptimal.

Furthermore, poor electromechanical coupling between the implanted patch and host myocardium may lead to arrhythmias. Nutrient diffusion also limits the thickness of patches, and currently, most designs are restricted to small animal models. Achieving clinically relevant, vascularized patch sizes remains a critical challenge that requires further investigation[56].

Integration of the Epicardium and Gene Therapy

Gene therapy involves the introduction of exogenous genes into cells *in vivo* or *in vitro* to achieve biological or therapeutic purposes. XC001, a novel adenovirus type-5 vector, is administered via a minimally invasive epicardial approach for the treatment of refractory angina. Volunteer trials have shown that XC001-VEGF gene therapy significantly improves both objective and subjective symptoms and has good tolerability[57]. However, research utilizing the epicardium as a gene delivery vector for gene therapy has yet to be fully explored[58].

Epicardial gene therapy involves brushing a solution containing a polymer mixture, vector, and diluted protease onto the atrial epicardium. The protease facilitates vector penetration, thereby enabling widespread atrial delivery while minimizing off-target effects[59]. Studies have demonstrated that this approach can deliver high-density adenoviral (Ad) vectors to atrial myocardium in pigs without evidence of off-target expression in organs[60-62]. This method circumvents blood and anatomical barriers, allowing the vector to reach all parts of the atrium via the epicardium. Clinically, this delivery strategy can be employed during open-heart surgery or heart transplantation and, applied modifications, could also be using thoracoscopy or catheter-based techniques[63]. The SmartSleeve enhances the effectiveness of therapy by allowing suture-free, conformal adhesion to the enabling epicardial surface and controlled, on-demand drug delivery in response electrocardiographic signals[64].

The CRISPR/Cas9 system, which uses a 20-nucleotide guide RNA, has emerged as a powerful gene-editing tool with the potential to correct genetic mutations[65, 66]. By using epicardial magnets for site-specific application, the CRISPR/Cas9 system can be locally delivered to the heart, reducing off-target effects in other tissues and cells, and controlling the duration of gene editing at specific *in vivo* sites to maximize therapeutic efficacy. In non-human primate models, the development of stronger externally actuated CRISPR/Cas9 magnetic complexes and further advancement of CRISPR/Cas9-mediated

targeted tissue therapeutic gene editing techniques will offer new avenues for cardiovascular disease treatment[67]. Wang et al. combined a functionalized self-assembling peptide (SAP) RADA16-I (RADA-RPR) with T\u03b4, enabling its adhesion to The functionalized SAP-released EPDCs. effectively activated the epicardium, induced EPDCs differentiation into cardiovascular and lymphatic endothelial cells, and promoted cardiomyocyte proliferation. After four weeks of treatment in a mouse myocardial infarction model, adverse cardiac remodeling was significantly attenuated and cardiac function was markedly improved[68].

In summary, existing studies have shown that EPDCs have achieved some results in improving cardiac function in rodents and hold early promise, but the administration of progenitor cells has not yet been translated into clinical practice. Their therapeutic effect is mainly mediated through secretory functions; however, the current cell retention rate is low. Therefore, it is necessary to improve the retention rate of therapeutic cells and enhance the release of key target factors[69]. In the treatment of myocardial injury, it is essential not only to focus on the delivery of exogenous cells but also to activate endogenous cells to ultimately replace lost cells and achieve repair. This includes stimulating endothelial cells to promote vascularization and inducing the proliferation of resident cardiomyocytes[70]. Tβ4, which promotes EPDCs inward migration and differentiation into endothelial and smooth muscle cells, is currently undergoing a multicenter phase I clinical trial for the treatment of cardiovascular diseases[71, However, there is still a long way to go before it can be widely applied in clinical practice.

Challenges in the Application of the Epicardium for Heart Failure Treatment

Although research on the epicardium has opened up new avenues for the treatment of heart failure, its clinical translation remains in the early stages, with many aspects of the technology and underlying mechanisms still underdeveloped. To enable its widespread application in regenerative medicine, several key scientific and technical challenges must be systematically addressed (Table 2).

Difficulty in Epicardial Cell Expansion and Differentiation *In Vitro*

Epicardial cells face significant challenges in terms of expansion and differentiation in vitro, mainly due to their limited quantity and lifespan. Although human epicardium and EPDCs hold great therapeutic potential, their limited availability hinders the advancement of related research. While EPDCs exhibit some proliferative ability, they often lose their epithelial phenotype after several passages, meaning that cells from different sources or passages may experimental influence subsequent in vitro outcomes[39]. Studies have shown that FGF-10 can promote the expansion of neonatal epicardial cells, but it does not significantly enhance cardiac regeneration[73]. Furthermore, when primary epicardial cells are isolated from E11.5 mouse hearts tissue adhesion methods, they considerable genetic variability during passaging, particularly in the expression of EMT-related genes. The EpiSV40 cell line, transfected with SV40-LT, shows promising proliferation, migration, and differentiation potential in vitro and serves as a suitable model for epicardial research[74].

Table 2. Brief Table of Key Challenges in the Application of the Epicardium for Heart Failure Treatment

Challenges	Specific Problems	References
Difficulty in epicardial cell expansion and differentiation <i>in vitro</i>	 Epicardial cells have limited quantity and lifespan, hindering in vitro expansion Loss of epithelial phenotype after passaging Significant variability among cells from different sources or passages Existing proliferation factors (e.g., FGF-10) have limited regenerative effects; primary cells exhibit genetic variability during culture; available cell lines (e.g., EpiSV40) require further validation for clinical use 	[39, 73, 74]
Poor survival and functional maintenance post-cell transplantation	 Transplanted cells are susceptible to immune rejection and inflammatory damage Challenges exist in achieving effective electromechanical coupling between transplanted and host cardiomyocytes, increasing risk of arrhythmias Optimal type and source of cardiac stem cells are still undefined Ensuring enhanced cell survival and sustained functionality post-transplantation remains a critical challenge 	[75-78]
Issues of biocompatibility and degradability in biomaterials	 Current biomaterials lack sufficient mechanical strength to support damaged myocardium (healthy myocardium stiffness ~50 kPa, heart failure myocardium ~200-300 kPa Current biomaterials are only ~10-20 Pa) Slow gelation rate of hydrogels may lead to blood flow obstruction and tissue necrosis Biomaterials need appropriate biodegradability matched with myocardial repair process to prevent long-term retention and adverse reactions 	[79, 80]
Safety and efficacy concerns in gene therapy	 Viral or non-viral vectors in gene therapy carry risks of tumorigenicity and immunogenicity CRISPR/Cas9 gene-editing technology faces immune toxicity and off-target effects Current detection techniques (e.g., Digenome-seq, CIRCLE-seq) have limited precision, highlighting the need for reducing off-target effects and immune toxicity 	[80-83]

Post-transplantation Cell Survival and Functional Maintenance

Immune rejection and inflammatory responses are among the main challenges in cell transplantation therapy. Although Phase I clinical trials have demonstrated that autologous skeletal muscle-derived cardiomyocytes can improve cardiac function after myocardial infarction, they also increased the incidence of ventricular arrhythmias, likely due to the failure of transplanted cells to establish proper electromechanical coupling with host cardiomyocytes[75-78]. Currently, the optimal type and source of cardiac stem cells remain undefined, making it crucial to address how to enhance cell survival and maintain function post-transplantation while minimizing immune rejection—a central issue in cell-based therapies for heart failure.

Issues of Biocompatibility and Degradability in Biomaterials

When using cellular or acellular biomaterials for cardiac repair, biocompatibility and mechanical properties are critical. Existing biomaterials often exhibit insufficient mechanical strength, making them incapable of providing adequate support to the damaged myocardium. For instance, biomaterials have stiffness levels ranging from 10 to 20 Pa, whereas the stiffness of a healthy heart is approximately 50 kPa and that of a heart with congestive heart failure reaches 200-300 kPa. As a result, these materials are too soft during end-diastole to support the infarcted region effectively[77]. Moreover, biomaterials must be biodegradable to allow gradual degradation as myocardial function recovers and remodeling occurs, preventing long-term retention and potential adverse reactions. However, the gelation speed of many biomaterials is relatively slow, which could block blood flow and lead to tissue necrosis[79].

Although cardiac patches have shown some progress, their clinical application is still limited. One key issue is the inability of the patches to achieve effective mechanical-electrical coupling with the host tissue, which can lead to arrhythmias. Therefore, developing biomaterials with improved mechanical strength, biocompatibility, and degradability remains a research priority.

Safety and Efficacy of Gene Therapy

Gene therapy involves introducing exogenous genes into cells either in vivo or in vitro to treat or prevent disease. However, safety concerns remain a significant hurdle. The use of viral and non-viral vectors requires thorough safety assessments before clinical translation[80]. Although viral vectors such as adenoviruses and herpesviruses have been employed in gene therapy, they pose risks of tumorigenesis and immunogenicity, which need to be addressed[81, 82]. Additionally, while the CRISPR/Cas9 system holds great potential for correcting genetic mutations, it may trigger immune toxicity and off-target effects. Techniques such as Digenome-seq and CIRCLE-seq show promise but still suffer from limitations in detection accuracy and read length[83]. Therefore, minimizing off-target effects and immune toxicity will be critical for the safe application of gene-editing technologies in the future. Scientific and regulatory barriers to gene therapy development exist in many countries, limiting global market expansion. In addition, the limited capacity of existing good manufacturing practice (GMP) facilities and the long production cycles for clinical materials further increase time and costs[84].

Future Perspectives

In the study of epicardial therapy for heart failure, future innovations are likely to focus on the following key areas (Table 3):

Table 3. A brief summary of future directions for epicardial therapy in heart failure

Future Directions	Potential Strategies and Innovations	References
Enhancing Expansion and Differentiation of EPDCs	Optimize culture conditions and signaling pathways (Wnt/ β -catenin, TGF- β) Employ SB431542 to maintain epithelial phenotype Co-culture EPDCs with cardiac progenitor cells (CMPCs) to enhance proliferation and regenerative capabilities	[85, 86]
Development of Novel Biomaterials and Scaffolds	Create elastic, biodegradable scaffolds mimicking cardiac mechanics Apply advanced 3D printing techniques for cardiac patch fabrication Integrate epicardium-derived ECM components (e.g., hyaluronic acid, fibronectin, type XII collagen) into biomaterial designs	[87]
Optimization of Gene Therapy Technologies	Refine CRISPR/Cas9 methods to reduce off-target effects Develop non-viral delivery systems (nanoparticles, liposomes) Utilize microRNAs and AAV vectors for targeted gene delivery and cardiac reprogramming	[88]
Combining Immunomodulation with Cardiac Repair	Exploit immunomodulatory effects of EPDCs-secreted cytokines Utilize Hippo pathway (YAP/TAZ) to recruit regulatory T cells, mitigating inflammation and adverse remodeling	[89]
Personalized Regenerative Strategies for Heart Failure	Tailor EPDCs and gene therapy approaches to individual genetic profiles and immune status Deliver Angpt4 protein precisely via nanoparticles for controlled regenerative responses Utilize patient-derived epicardial-like cells from induced pluripotent stem cells	[90-92]

Enhancing the Expansion and Differentiation Capacity of Epicardium-Derived Cells

EPDCs have very limited ability to expand and differentiate in vitro. Future studies should aim to develop more efficient culture methods to overcome the loss of epithelial phenotype and multipotency during in vitro proliferation. Optimizing culture conditions, applying specific growth factors and cytokines, or regulating key signaling pathways such as Wnt/ β -catenin and TGF- β through gene editing techniques may enhance EPDCs proliferation and differentiation, thereby improving their reparative capacity. Researchers may also explore combining EPDCs with other stem cell types to enhance their plasticity and regenerative potential. For example, culturing EPDCs in media containing SB431542, an ALK5 (TGF-β type I receptor) inhibitor, helps maintain their epithelial phenotype and allows for monolayer expansion[86]. In addition, co-culturing EPDCs with cardiac progenitor cells (CMPCs) isolated from human hearts has been shown to enhance EPDCs proliferation, as demonstrated by Ki67 analysis. When co-transplanted into infarcted mouse hearts, the combined cells exhibited a superior effect on cardiac remodeling through the secretion of matrix-modulating factors, showing synergistic benefits compared to single-cell-type therapies[85].

Development of Novel Biomaterials and Scaffolds

Current biomaterials and scaffolds for cardiac repair often lack sufficient mechanical strength and biocompatibility. Future efforts should focus on developing more elastic and biodegradable scaffolds that better mimic the mechanical properties of native cardiac tissue. Advances in 3D printing offer new possibilities for the precise design of biomaterials. This technology can be used to fabricate cardiac patches with optimized geometry, pore size, and diameter, thereby improving nutrient and oxygen transport, promoting cell proliferation, and enhancing cardiac function. Epicardium-derived ECM materials are also expected to play a key role in supporting cell proliferation, migration, and differentiation. Future research should explore how integrate to epicardium-derived ECM with biomaterials to enhance cardiac repair. For instance, the epicardium secretes hyaluronic acid (HA), which swells upon hydration and modulates epicardial cell EMT, migration, angiogenesis, myocardial and regeneration. After cardiac injury, integrin receptors on cardiomyocytes bind to fibronectin secreted by epicardial cells, enhancing regenerative signaling. Additionally, the marked upregulation of type XII

collagen in the epicardium post-injury is closely linked to myocardial repair[87].

Further Optimization of Gene Therapy Technologies

As an innovative therapeutic strategy, gene therapy holds promise for breakthroughs in heart failure treatment. Advances in CRISPR/Cas9 technology have opened new possibilities for gene editing in EPDCs. Future studies should focus on optimizing the CRISPR/Cas9 system to minimize off-target effects while improving specificity and safety. Additionally, the development of novel non-viral vectors-such as nanoparticles and liposomes - will be key to enhancing delivery efficiency and targeting. Non-coding microRNAs (miRNAs), which can simultaneously target multiple mRNAs. have the potential to reprogram cardiomyocytes into proliferative Adeno-associated viruses (AAVs) can selectively target cardiomyocytes, reducing the risk of ectopic gene expression in non-cardiac cells[88]. Integrating EPDCs with gene therapy may allow precise regulation of cardiac repair and facilitate functional recovery.

Combining Immunomodulation with Cardiac Repair

Epicardial cells not only contribute to structural cardiac repair but also possess immunomodulatory capabilities that help reduce inflammation and adverse remodeling. Future research may focus on leveraging EPDCs to modulate immune responses and reduce rejection following heart or cell transplantation. Inducing EPDCs to secrete specific cytokines or exploiting their immunomodulatory functions could suppress pathological remodeling, improve cardiac function, and enhance clinical outcomes. For example, components of the epicardial Hippo signaling pathway, YAP and TAZ, can recruit regulatory T cells (Tregs) to the injured myocardium, suppressing post-infarction inflammation limiting adverse remodeling[89].

Personalized Regenerative Strategies for Heart Failure

The future application of the epicardium in heart failure therapy is expected to move toward personalized medicine. With the advancement of precision medicine, treatment strategies can be tailored to individual patients based on specific disease conditions and the type of cardiac injury. For instance, in patients with heart failure with preserved ejection fraction (HFpEF), combining EPDCs with targeted gene therapy may help restore cardiac

function. Developing personalized stem cell and gene therapy approaches based on individual immune status and genetic background could improve therapeutic efficacy and reduce side effects. For example, following cardiac injury, EPDCs can secrete Angpt4, which activates the Tie2-MAPK pathway in endocardial cells to promote regeneration. As a secreted protein, Angpt4 is easy to deliver and allows precise control over dosage and duration compared to intracellular proteins[90]. Nanoparticles, capable of carrying multiple targeted molecules, offer flexibility in drug delivery and may pave the way for personalized medicine[91]. Additionally, systems have been developed to maintain the epithelial phenotype of adult human epicardial cells in vitro, and methods exist to induce pluripotent stem cells to differentiate into epicardial-like cells via Wnt signaling modulation. These patient-derived cell types offer a feasible path toward individualized cardiac therapy[92].

Summary

The epicardium and EPDCs, with their intrinsic potential to differentiate into multiple cardiac lineages-including cardiomyocytes-have emerged as a promising cell source for cardiac regenerative therapy. Their key roles in subepicardial matrix remodeling and paracrine signaling further enhance their therapeutic appeal. Increasing evidence supports the involvement of the epicardium and EPDCs in heart failure treatment. However, challenges remain in maintaining their phenotypic stability multipotency during in vitro expansion, as well as ensuring the safety, efficacy, and personalization of gene therapies. Future research should focus on harnessing the multipotent capacity of EPDCs by identifying optimal stimulatory factors, integrating gene editing technologies, and combining them with advanced biomaterials to enhance their regenerative potential and promote differentiation into functional cardiomyocytes. Personalized treatment strategies tailored to individual patient profiles may further improve therapeutic outcomes and provide strong support for the advancement of heart failure therapies.

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Author Contributions

Chenlei Zhou and Yaping Xu wrote the first draft of the manuscript. Zhikun Guo revised the manuscript. All authors critically reviewed the article and gave final approval of the manuscript.

Data Availability

All data generated or analyzed in this study are included in this published article. Any other data will be available on reasonable request from corresponding author.

Competing Interests

The authors have declared that no competing interest exists.

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