SUPPLEMENTARY MATERIALS

Aortic calcification accelerates cardiac dysfunction via inducing apoptosis of cardiomyocytes

Nannan Hao^{1, *}, Hui Yong^{1, *}, Feifei Zhang^{1, *}, Chang Liu², Yulu Qiu², Yumeng Shi²,

Chunjian Li^{1, ‡}, Fang Wang^{1, ‡}

¹Department of Cardiology, the First Affiliated Hospital of Nanjing Medical University, China

²Department of Rheumatology, the First Affiliated Hospital of Nanjing Medical University, China

*These authors contributed equally to this work.

[†]Correspondence to: Dr. Fang Wang, E-mail: <u>wangfangheart@njmu.edu.cn</u>, or Chunjian Li, <u>lijay@njmu.edu.cn</u>, Department of Cardiology, the First Affiliated Hospital of Nanjing Medical University, Nanjing 210029, China.

Supplementary methods

Patient blood samples

Blood samples were collected from peripheral veins and centrifuged to obtain plasma from 3 CAD patients (aged 67 ± 2.65 years, 2 males and 1 female), 3 CKD patients (aged 54 ± 4.51 years, 2 males and 1 female) and 3 healthy controls (aged 36 \pm 3.71 years, 2 males and 1 female). All patients underwent coronary computed tomography angiogram (CTA) with coronary artery calcification (Agatston score > 400). This study was approved by the ethics committee of the First Affiliated Hospital of Nanjing Medical University.

Plasma from patients treated H9C2 cells

To study effect of peripheral blood from patients with vascular calcification (VC) on cardiomyocytes function, we cultured H9C2 cells with plasma collected from both CAD and CKD patients with coronary artery calcification as well as healthy controls at different dilutions with FBS. After 48 h, H9C2 cells were collected for RT-qPCR analysis (ANP, BNP and TGF- β) and TUNEL staining, respectively.

Von Kossa staining

Von Kossa staining was used for calcification determination. The paraffin-embedded heart and kidney from mice were stained with a von Kossa staining kit according to the manufacturer's protocol. Briefly, after rinsing in ddH2O, sections were incubated with 1% silver nitrate solution under a 60 watt light bulb for 1 hour. After washing, 5% sodium thiosulfate was added to the slides for 5 min to remove un-reacted silver.

Calcified areas are shown as black or brown black deposits.

Supplementary figures



Supplementary Figure 1. The kidney changes of VD₃-induced VC mice. (A) C57BL/6 mice were intraperitoneally injected with VD₃ (350,000 IU/kg/day) for 14 days. At day 42, kidney samples were collected and the ratio of kidney to body weight was determined. Values are shown as mean \pm SEM (NC = 5, VD = 8). **P < 0.01. (B) Representative images of H&E, Von kossa and ARS staining for mice kidney, respectively. VC: vascular calcification; ARS: Alizarin Red S.



Supplementary Figure 2. Plasma from CAD or CKD patients induces apoptosis of rat cardiomyocytes. (A) RT-qPCR analysis of the expression of cardiac dysfunction related genes (ANP, BNP and TGF- β) in H9C2 cells under different plasma from patients with CAD, CKD and healthy controls (HC) for 48 h. (B-C) Representative images and quantification of TUNEL staining for H9C2 cells apoptosis with different plasma from CAD patients and healthy controls (HC) (B) as well as CKD patients (C) for 48 h. Data from one representative experiment carried out in triplicate and shown as mean \pm SEM. *P < 0.05, **P < 0.05, **P < 0.001. CAD: coronary artery disease; CKD: chronic kidney disease; CAC: coronary artery calcification.

Supplementary tables

Gene symbol	Forward (5'-3')	Reverse (5'-3')
mouse ANP	AGGCAGTCGATTCTGCTT	CGTGATAGATGAAGGCAGGAAG
mouse BNP	TAGCCAGTCTCCAGAGCAATTC	TTGGTCCTTCAAGAGCTGTCTC
mouse MMP2	CAAGTTCCCCGGCGATGTC	TTCTGGTCAAGGTCACCTGTC
mouse MMP9	GACGACATAGACGGCATCC	TGGTTCAGTTGTGGTGGTG
mouse TGF-β	CTCCCGTGGCTTCTAGTGC	GCCTTAGTTTGGACAGGATCTG
mouse β-actin	CCTCACTGTCCACCTTCC	GGGTGTAAAACGCAGCTC
rat ANP	GCCGGTAGAAGATGAGGTCA	GGGCTCCAATCCTGTCAATC
rat BNP	ATCTGTCGCCGCTGGGAGGT	TGGATCCGGAAGGCGCTGTC
rat TNFα	CAGCCAGGAGGGAGAAC	GTATGAGAGGGACGGAACC
rat IL-6	CCGTTTCTACCTGGAGTTTGT	GTTTGCCGAGTAGACCTCATAG
rat MMP2	TTGACCAGAACACCATCG	CTTGCGGGGAAAGAAGT
rat MMP9	CGCTGGGCTTAGATCATT	TGCTGGATGCCTTTTATGT
rat TGF-β	CCTACATTTGGAGCCTGGA	CCGGGTTGTGTTGGTTG
rat Colla1	GAGCGGAGAGTACTGGATCGA	CTGACCTGTCTCCATGTTGCA
rat Col3a1	TGCCATTGCTGGAGTTGGA	GAAGACATGATCTCCTCAGTGTTGA
rat β-actin	CCCGCGAGTACAACCTTCT	CGTCATCCATGGCGAACT

Table 1 Primers used for quantitative real time PCR

Indicators	NC group (n = 5)	VD group (n = 8)	
Body weight (g)	25.03 ± 0.55	$15.21 \pm 0.43 **$	
Heart weight (g)	0.12 ± 0.002	0.08 ± 0.002 **	
Liver weight (g)	1.24 ± 0.069	0.78 ± 0.06 **	
kidney weight (g)	0.36 ± 0.009	0.39 ± 0.07	

Table 2 Body and organ weights of normal and VD₃-induced VC mice

Data are presented as mean \pm SEM. NC: normal control; VD: VD₃-treated mice. **P <

0.01.

Table 3 Echocardiographic data of left ventricle in normal and VD₃-induced VC

Indicators	NC (n = 5)	VD (n = 10)	P value
FS (%)	45.830 ± 1.711	41.380 ± 1.035	0.035*
LVEF (%)	77.620± 1.614	72.950 ± 1.064	0.044*
LVMI (g/m2)	15.710 ± 0.483	14.850 ± 0.403	0.218
Heart rate	595.00 ± 19.65	445.30 ± 19.24	0.0003***
LV Mass Cor	100.400 ± 3.327	71.90 ± 5.24	0.003*
Volume; d (µl)	50.720 ± 2.371	44.130 ± 1.908	0.059
Diameter; d (mm)	3.491 ± 0.067	3.292 ± 0.059	0.063
Diameter; s (mm)	1.895 ± 0.092	1.930 ± 0.051	0.723
LVAWT; s (mm)	1.639 ± 0.037	1.381 ± 0.049	0.005**
LVAWT; d (mm)	1.040 ± 0.033	0.909 ± 0.028	0.014*
LVPWT; d (mm)	0.916 ± 0.010	0.817 ± 0.018	0.003**
LVPWT; s (mm)	1.411 ± 0.016	1.245 ± 0.023	0.0004***
Stroke Volume (µl)	39.390± 1.183	32.340± 1.377	0.006**
Cardiac Output (mL/min)	23.420 ± 0.895	14.310 ± 0.643	< 0.0001***
PWTF (%)	58.030 ± 3.832	51.800± 2.227	0.156
CI	2949.0 ± 181.2	2166.0 ± 119.0	0.003**

mice

Data are presented as mean \pm SEM. *P < 0.05, **P < 0.01, ***P < 0.001. FS: fractional shortening; LVEF: left ventricular ejection fraction; LVMI: left ventricular mass index; d: diastolic diameter; s: systolic diameter; LVAWT: left ventricular anterolateral wall thickness; LVPWT: left ventricular posterior wall thickness; PWTF: posterior wall thickness; CI: cardiac index.