

Research Paper

Recipient Cytotoxic T Lymphocyte Antigen 4 +49 Single-Nucleotide Polymorphism Is Not Associated with Acute Rejection after Liver Transplantation in Chinese Population

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Abstract

Objective: Single-nucleotide polymorphisms (SNPs) in Cytotoxic T lymphocyte antigen 4 (CTLA-4) gene have been detected and proved to associate with the incidence of rejection after transplantation. However, previous studies gained inconsistent results about the association between CTLA-4 +49 single-nucleotide polymorphism and susceptibility of allograft rejection. Therefore we sought to clarify whether CTLA-4 +49 SNP influences the incidence of acute rejection after liver transplantation in Chinese population. **Methods:** Genomic DNA from 335 liver transplant recipients was genotyped for CTLA-4 +49 SNP by DNA sequencing. Acute rejection was confirmed by pathologic evidences. The association between CTLA-4 +49 SNP and incidence of acute rejection was then analyzed by dominant, recessive, codominant and overdominant models. **Results:** The incidence of acute rejection within the first 3 months was 11.9%. In acute rejectors, the frequency was 45% for G/G, 10% for A/A and 45% for A/G respectively, compared with 47.5% for G/G, 10.8% for A/A and 41.7% for A/G in non-acute rejectors. And no significant difference of allele distribution between these 2 groups was detected. **Conclusions:** This study suggests that CTLA-4 +49 SNP is not associated with acute rejection after liver transplantation in Chinese population.

Key words: Cytotoxic T lymphocyte antigen 4, liver transplantation, Single Nucleotide Polymorphism.

Introduction

T cell activation plays a central role in initiating the cascade of rejection after liver transplantation. Two separate signals are required for T cell activation: the antigen-specific signals through T-cell receptor (TCR) and the costimulatory signals from the interaction between CD28 and B7-1, B7-2 [1]. In recent years, costimulatory pathway has gained increasing interest in transplant immunology. Many reports indicated

that blocking of costimulatory pathway can prevent transplant rejection and even induce long-term allograft tolerance [2-4]. CTLA-4, which shares the same ligands with CD28, is expressed on activated T cells and provides an inhibitory signal in immune responses [5, 6]. And it is believed to play a key role in inducing peripheral tolerance [7, 8]. Several mechanisms are demonstrated to explain the T cell inhibi-

tion caused by engagement of CTLA-4: the competitive antagonist of CD28 for B7 as well as the direct inhibitory signals mediated by CTLA-4 cytoplasmic tail [9]. Moreover, CTLA-4 is also expressed on CD4⁺CD25⁺ regulatory T cell, and is believed to play a key role in controlling regulatory T cell activation and therefore contribute to the development of transplant tolerance [10, 11]. Recently CTLA-4Ig has been developed to competitively inhibit the interaction between CD28 and B7 molecules and is proved to prolong the allograft survival, which not only highlights the importance of CTLA-4 in transplant immunology but also offers the potential for more immune-selective immunosuppressive agents [12-14].

CTLA-4 +49 is one of the most well-studied polymorphisms in CTLA-4 gene and is generally considered to play an important role in the individual difference of immune intensity mediated by costimulatory pathway [15]. The A to G variation at position +49 in exon 1 leads to a threonine to alanine substitution at position 17 of the signal peptide of the CTLA-4 molecule. CTLA-4 +49 G homozygote leads to decreased CTLA-4 surface expression [16-18] and is believed to reduce the inhibitory function of CTLA-4 on T cell proliferation [18, 19]. The G allele is reported to associate with many autoimmune diseases, as type 1 autoimmune hepatitis, primary biliary cirrhosis [20], autoimmune thyroid disease [21], insulin-dependent diabetes mellitus [22], and rheumatoid arthritis [23]. Moreover, recent studies also reported that CTLA-4 +49 SNP is associated with the outcome after kidney transplantation [24]. However, the available reports about association between CTLA-4 +49 SNP and the susceptibility of allograft rejection are inconsistent [25-28]. The association between CTLA-4 +49 SNP and acute rejection after liver transplantation in Han Chinese population are rarely studied. Therefore, in the view of the importance of costimulatory pathway in the transplant immunology as well as the limited and inconclusive results about the role of CTLA-4 +49 SNP in acute allograft rejection in China, we examined and tried to clarify the possible involvement of CTLA-4 +49 SNP in acute rejection after liver transplantation in Han Chinese population.

Patients and methods

Patients enrollment

This study included 335 patients who underwent orthotopic liver transplantation in the Division of Hepatobiliary Pancreatic Surgery, Department of Surgery, First Affiliated Hospital, School of Medicine, Zhejiang University, from 2005 to 2010. The age of patients ranges from 13 to 71 years old. There were

290 male and 45 female involved. And all patients included survived longer than 6 months after transplantation. Written informed consent was obtained from all participants or their guardians and the study protocol was approved by the Ethics Committee of our hospital. The protocol conforms with the ethical guidelines of the 1975 Helsinki Declaration. Routine immunosuppression regimen consisted of tacrolimus or Rapamycin or Cyclosporine with mycophenolate mofetil and steroids. Patients showing clinical symptoms of acute rejection within the first 6 months after liver transplantation underwent liver biopsy. And acute rejection was confirmed by pathologic evidences according to Banff criteria. Treatments of prednisone were given to patients having acute rejection. Those did not have any rejection episode were considered as nonrejecors.

CTLA-4 genotyping

DNA was isolated from peripheral blood mononuclear cell of the recipients using the QIAamp DNA blood mini kit (Qiagen Inc. Hilden, Germany). Recipient CTLA-4 +49 SNP was typed by means of DNA sequencing. PCR reaction system (10 μ l) included 1x GC buffer (TAKARA, China), 3.0mM Mg²⁺, 0.3mM dNTP, 1 U HotStarTaq polymerase (Qiagen Inc. Hilden, Germany), 1 μ l templates DNA and 1 μ l PCR primers: rs231775F:CCATGGCTTGCCTTGGATTTC, rs231775R: CCAGCCAAGCCAGATTGGAGTT. The cycling program was as follows: denaturation at 95°C for 2 min followed by 11 cycles x (94°C 20s, 65°C-0.5°C /cycle 40s, 72°C 1min 30s); 24 cycles x (94°C 20s, 59°C 30s, 72°C 1.5min); 72°C 2 min. Thereafter, 1U SAP and 1U Exonuclease I were added to 10 μ L of PCR product for purification. The mixture was incubated at 37°C for 60min, followed by incubation at 75°C for 15min. Then, 2 μ L purified PCR product was mixed with 5 μ l SNaPshot Multiplex Kit (ABI, USA), 2 μ l Ultrapure water and 1 μ l sequencing primer, which is rs231775SR: TTTTTTTTTTTTTTGTGCAGGGCCA GGTCTGG, for further reaction. The cycling program was as follows: 96°C 1min; 28 x (96°C 10s, 50°C 5s, 60°C 30s). Then, 1U SAP was added to 10 μ L of PCR product for purification. The mixture was incubated at 37°C for 60min, followed by incubation at 75°C for 15 min. Then, 0.5 μ L purified PCR product was mixed with 0.5 μ l Liz120 SIZE STANDARD and 9 μ l Hi-Di, and then incubated at 95 °C for 5 min before loaded onto an ABI3730XL Genetic Analyzer (ABI, USA). And the data was analyzed using GeneMapper 4.0 (AppliedBiosystems, USA).

Statistical analysis

The SNP data were evaluated for Har-

dy-Weinberg equilibrium (bioinfo.iconcologia.net/snpstats/start.htm). Differences in the characteristics of recipients between the rejection group and the nonrejection group were analyzed by chi-square test, using SPSS software for Windows (Statistical Product and Service Solutions, version 14.0, SSPS Inc, Chicago, IL, USA). CTLA-4 genotypes were analyzed by SNPstats (bioinfo.iconcologia.net/snpstats/start.htm). $P < 0.05$ was considered statistical significance.

Results

The incidence of acute rejection within the first 6 months was 40 of 335 (11.9%). The primary diseases in this study were hepatocellular carcinoma (HCC) (37.3%), fulminant hepatitis (17.0%), alcoholic cirrhosis (4.8%) and others (40.9%). There was no significant

association between primary diseases and incidence of acute rejection ($P > 0.05$). Neither the recipient age, gender nor blood type was significantly different between rejectors and nonrejectors ($P > 0.05$).

We genotyped CTLA-4 +49 SNP in this study and the results were showed in Table 1. The CTLA-4 +49 SNP was in Hardy-Weinberg equilibrium in the cohort. The frequency of the G allele was 68.2% among all patients. In acute rejectors, the frequency was 45% for G/G, 10% for A/A and 45% for A/G respectively, compared with 47.5% for G/G, 10.8% for A/A and 41.7% for A/G in non rejectors. And none of the dominant, recessive, codominant or overdominant models revealed any significant association between CTLA-4 +49 SNP and incidence of acute rejection ($P > 0.05$).

Table 1. CTLA-4 +49 snp in liver transplantation recipients in relation to acute rejection.

Model	Genotype	NAR (n=295)	AR (n=40)	OR(95% CI)	P-value
Codominant	G/G	140 (47.5%)	18(45%)	1.00	0.92
	G/A	123 (41.7%)	18(45%)	1.10(0.55-2.22)	
	A/A	32 (10.8%)	4(10%)	0.96(0.30-3.03)	
Dominant	G/G	140 (47.5%)	18(45%)	1.00	0.77
	G/A-A/A	155 (52.5%)	22(55%)	1.07(0.55-2.09)	
Recessive	G/G-A/A	263 (89.2%)	36(90%)	1.00	0.87
	A/A	32 (10.8%)	4(10%)	0.91(0.31-2.73)	
Overdominant	G/G-A/A	172 (58.3%)	22(55%)	1.00	0.69
	G/A	123 (41.7%)	18(45%)	1.11(0.57-2.16)	

AR, acute rejectors; NAR, non-acute rejectors.

Discussion

CTLA-4, one of the most extensively investigated costimulatory molecule, acts as a negative regulator of peripheral T cell function. The importance of CTLA-4 in autoimmune diseases, antitumor immunology and transplant immunology has been demonstrated and emphasized [2, 9, 29]. Polymorphisms in CTLA-4 influence the expression, transcription and translation of CTLA-4 and therefore affect its function. Particular polymorphisms such as SNPs in position +49, +106 and -1722 have been reported to influence the susceptibility to autoimmune diseases as Graves' disease, autoimmune hypothyroidism, multiple sclerosis and systemic lupus erythematosus [16, 19, 30, 31]. And high incidence of CTLA-4 AA(CT60) polymorphism has been observed in renal cell cancer [32]. CTLA-4 +49 SNP was also reported to play an important role in non-Hodgkin's lymphoma and oral squamous cell carcinoma [33, 34]. Moreover, the DNA variants of CTLA-4, such as SNPs in +49, -318 and the dinucleo-

tide (AT)_n repeat polymorphism, are involved in the outcome of solid organ transplantation [24, 28, 35].

CTLA-4 gene encodes for 2 isoforms of CTLA-4: a membrane bound CTLA-4 (mCTLA-4) and a soluble CTLA-4 (sCTLA-4) [36]. The CTLA-4 +49 A/G polymorphism leads to a threonine to alanine change in amino acid 17 of CTLA-4, which decreases the cell surface expression of mCTLA-4. As a result, CTLA-4 +49 G/G exerts diminished inhibitory effects on T cell proliferation [18, 19]. The protective effect of CTLA-4 +49 A allele in autoimmune diseases has been identified by previous studies. In our study, we enrolled 335 patients who underwent liver transplantation and used dominant, recessive, codominant and overdominant models to evaluate the association between the A/G polymorphism and acute allograft rejection. We found that the allele frequencies in recipients were consistent with normal Chinese population [37]. No significant association between CTLA-4 +49 SNP and incidence of acute rejection has been detected, which was consistent with the results of Slavcheva et al [28].

The continuous use of immunosuppression might be a reasonable explanation for the different effects CTLA-4 +49 A allele has in acute rejection and auto-immune diseases. mCTLA-4 is expressed only after T cell activation. Both CD28 and IL-2 play important roles in the up-regulation of CTLA-4 expression. The surface expression of CTLA-4 decreases significantly when IL-2 production is blocked using cyclosporin A [38]. Therefore it is conceivable that immunosuppressive agencies as tacrolimus can inhibit the expression of mCTLA-4 by inhibiting the activation of T cells and the production of IL-2. As a result, CTLA-4 +49 A allele has decreased or even no protective effect against acute rejection under the immunosuppressive therapies.

The studies of Marder et al showed that recipients with CTLA4 +49 G/G genotype had decreased graft survival time compared with A/A and A/G [27]. However the main cause of graft failure is the recurrence of HCV rather than acute rejection. Thus the results of Marder and colleagues are not contradicted with ours. Kwekkeboom and colleagues found that recipient CTLA-4 +49 G/G genotype is associated with reduced acute rejection after liver transplantation [25]. Similarly, Karimi et al also found increased A allele prevalence in rejectors after liver transplantation in Iranian patients [39]. Such results seemed to be unexplainable by previous studies. But the further studies of Kwekkeboom et al soon offered a possible explanation: the CTLA-4 +49 A/+6230 G haplotype is a co-dominant risk allele for acute rejection after liver transplantation. The CTLA-4 +6230 SNP was reported to influence the sCTLA-4 production [26]. The sCTLA-4 is produced by nonactivated T cells and could either inhibit or exacerbate the immune response: during the initiation, sCTLA-4 can block the interaction of CD28-B7; and at latter stages, it blocks the CTLA-4-B7 interaction [36]. Therefore, The CTLA-4 +6230 SNP may influence the incidence of acute rejection by sCTLA-4 secretion. However, the reason why the risk allele is +49A+6230G haplotype, rather than +49G+6230G remains unclear. One possible explanation is that another unknown single allele is linked to the +49A+6230G haplotype and inhibits CTLA-4 function. Moreover, previous research showed that, several SNPs in CTLA-4 gene are related to the outcome of transplantation. It is possible that multiple alleles or genes contribute the susceptibility to acute rejection after liver transplantation and CTLA-4 +49 SNP does not act as primary susceptibility polymorphism. And the inconsistent results may be attributed to different genetic background of different ethnicities, small study number and baseline statistical significance. More studies are needed to

demonstrate these hypotheses.

In conclusion, our study reveals that CTLA-4 +49 SNP has no significant association with acute rejection after liver transplantation in Chinese population. Until now, CTLA4 +49 SNP alone is not a promising predictor for the incidence of acute rejection after liver transplantation.

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Competing Interests

The authors have declared that no competing interest exists.

References

1. Turka LA, Linsley PS, Lin H, Brady W, Leiden JM, Wei RQ, et al. T-cell activation by the CD28 ligand-B7 is required for cardiac allograft-rejection *in vivo*. *Proc Natl Acad Sci U S A*. 1992; 89: 11102-5.
2. Zheng XG, Turka LA. Blocking T-cell costimulation to prevent transplant rejection. *Transplantation Proceedings*. 1998; 30: 2146-9.
3. Azuma H, Chandraker A, Nadeau K, Hancock WW, Carpenter CB, Tilney NL, et al. Blockade of T-cell costimulation prevents development of experimental chronic renal allograft rejection. *Proc Natl Acad Sci U S A*. 1996; 93: 12439-44.
4. Schaub M, Stadlbauer THW, Chandraker A, Vella JP, Turka LA, Sayegh MH. Comparative strategies to induce long-term graft acceptance in fully allogeneic renal versus cardiac allograft models by CD28-B7 T cell costimulatory blockade: Role of thymus and spleen. *J Am Soc Nephrol*. 1998; 9: 891-8.
5. Krummel MF, Allison JP. CD28 and CTLA-4 have opposing effects on the response of T-cells to stimulation. *Journal of Experimental Medicine*. 1995; 182: 459-65.
6. Walunas TL, Lenschow DJ, Bakker CY, Linsley PS, Freeman GJ, Green JM, et al. CTLA-4 can function as a negative regulator of T-cell activation. *Immunity*. 1994; 1: 405-13.
7. Perez VL, VanParijs L, Biuckians A, Zheng XX, Strom TB, Abbas AK. Induction of peripheral T cell tolerance *in vivo* requires CTLA-4 engagement. *Immunity*. 1997; 6: 411-7.
8. Tivol EA, Borriello F, Schweitzer AN, Lynch WP, Bluestone JA, Sharpe AH. Loss of CTLA-4 leads to massive lymphoproliferation and fatal multiorgan tissue destruction, revealing a critical negative regulatory role of CTLA-4. *Immunity*. 1995; 3: 541-7.
9. Hodi FS. Cytotoxic T-Lymphocyte - Associated antigen-4. *Clinical Cancer Research*. 2007; 13: 5238-42.
10. Kingsley CI, Karim M, Bushell AR, Wood KJ. CD25(+)/CD4(+) regulatory T cells prevent graft rejection: CTLA-4- and IL-10-dependent immunoregulation of alloresponses. *Journal of Immunology*. 2002; 168: 1080-6.
11. Okumura A, Ishikawa T, Sato S, Yamauchi T, Oshima H, Ohashi T, et al. Deficiency of forkhead box P3 and cytotoxic T-lymphocyte-associated antigen-4 gene expressions and impaired suppressor function of CD4(+)/CD25(+) T cells in patients with autoimmune hepatitis. *Hepatology Research*. 2008; 38: 896-903.
12. Kirk AD, Harlan DM, Armstrong NN, Davis TA, Dong YC, Gray GS, et al. CTLA4-Ig and anti-CD40 ligand prevent renal allograft rejection in primates. *Proc Natl Acad Sci U S A*. 1997; 94: 8789-94.
13. Larsen CP, Pearson TC, Adams AB, Tso P, Shirasugi N, Strobert E, et al. Rational development of LEA29Y (belatacept), a high-affinity variant of CTLA4-Ig with potent immunosuppressive properties. *American Journal of Transplantation*. 2005; 5: 443-53.
14. Vincenti F, Muehlbacher F, Nashan B, Larsen C, Atillasoy E, Natarajan K, et al. Co-stimulation blockade with LEA29Y in a calcineurin inhibitor

- free maintenance regimen in renal transplant: 6-month efficacy and safety. *American Journal of Transplantation*. 2004; 4: 442.
15. Ueda H, Howson JMM, Esposito L, Heward J, Snook H, Chamberlain G, et al. Association of the T-cell regulatory gene CTLA4 with susceptibility to autoimmune disease. *Nature*. 2003; 423: 506-11.
 16. Ligers A, Teleshova N, Masterman T, Huang WX, Hillert J. CTLA-4 gene expression is influenced by promoter and exon 1 polymorphisms. *Genes Immun*. 2001; 2: 145-52.
 17. Anjos S, Nguyen A, Ounissi-Benkhalha H, Tessier MC, Polychronakos C. A common autoimmunity predisposing signal peptide variant of the cytotoxic T-lymphocyte antigen 4 results in inefficient glycosylation of the susceptibility allele. *Journal of Biological Chemistry*. 2002; 277: 46478-86.
 18. Maurer M, Loserth S, Kolb-Maurer A, Ponath A, Wiese S, Kruse N, et al. A polymorphism in the human cytotoxic T-lymphocyte antigen 4 (CTLA4) gene (exon 1+49) alters T-cell activation. *Immunogenetics*. 2002; 54: 1-8.
 19. Kouki T, Sawai P, Gardine CA, Fisfalen ME, Alegre ML, DeGroot LJ. CTLA-4 gene polymorphism at position 49 in exon 1 reduces the inhibitory function of CTLA-4 and contributes to the pathogenesis of Graves' disease. *Journal of Immunology*. 2000; 165: 6606-11.
 20. Miyake Y, Ikeda F, Takaki A, Nouse K, Yamamoto K. +49A/G polymorphism of cytotoxic T-lymphocyte antigen 4 gene in type 1 autoimmune hepatitis and primary biliary cirrhosis: A meta-analysis. *Hepatology Research*. 2011; 41: 151-9.
 21. Kavvoura FK, Akamizu T, Awata T, Ban Y, Chistiakov DA, Frydecka I, et al. Cytotoxic T-lymphocyte associated antigen 4 gene polymorphisms and autoimmune thyroid disease: A meta-analysis. *Journal of Clinical Endocrinology & Metabolism*. 2007; 92: 3162-70.
 22. Donner H, Rau H, Walfish PG, Braun J, Siegmund T, Finke R, et al. CTLA4 alanine-17 confers genetic susceptibility to Graves' disease and to type 1 diabetes mellitus. *Journal of Clinical Endocrinology & Metabolism*. 1997; 82: 143-6.
 23. Benhatchi K, Jochmanova I, Habalova V, Wagnerova H, Lazurova I. CTLA4 exon1 A49G polymorphism in Slovak patients with rheumatoid arthritis and Hashimoto thyroiditis-results and the review of the literature. *Clinical Rheumatology*. 2011; 30: 1319-24.
 24. Gorgi Y, Sfar I, Ben Abdallah T, Abderrahim E, Ayed SJ, Aouadi H, et al. CTLA-4 exon 1 (+49) and promoter (-318) gene polymorphisms in kidney transplantation. *Transplantation Proceedings*. 2006; 38: 2303-5.
 25. de Reuver P, Pravica V, Hop W, Boor P, Metselaar HJ, Hutchinson IV, et al. Recipient CTLA-4+49 G/G genotype is associated with reduced incidence of acute rejection after liver transplantation. *American Journal of Transplantation*. 2003; 3: 1587-94.
 26. Tapirdamaz O, Pravica V, Metselaar HJ, Hansen B, Moons L, van Meurs JBJ, et al. Polymorphisms in the T cell regulatory gene cytotoxic T lymphocyte antigen 4 influence the rate of acute rejection after liver transplantation. *Gut*. 2006; 55: 863-8.
 27. Marder BA, Schroppel B, Lin M, Schiano T, Parekh R, Tomer Y, et al. The impact of costimulatory molecule gene polymorphisms on clinical outcomes in liver transplantation. *American Journal of Transplantation*. 2003; 3: 424-31.
 28. Slavcheva E, Albanis E, Jiao QS, Tran H, Bodian C, Knight R, et al. Cytotoxic T-lymphocyte antigen 4 gene polymorphisms and susceptibility to acute allograft rejection. *Transplantation*. 2001; 72: 935-40.
 29. Luhder F, Hoglund P, Allison JP, Benoist C, Mathis D. Cytotoxic T lymphocyte-associated antigen 4 (CTLA-4) regulates the unfolding of autoimmune diabetes. *Journal of Experimental Medicine*. 1998; 187: 427-32.
 30. Hudson LL, Rocca K, Song YW, Pandey JP. CTLA-4 gene polymorphisms in systemic lupus erythematosus: a highly significant association with a determinant in the promoter region. *Human Genetics*. 2002; 111: 452-5.
 31. Kotsa K, Watson PF, Weetman AP. A CTLA-4 gene polymorphism is associated with both Graves' disease and autoimmune hypothyroidism. *Clinical Endocrinology*. 1997; 46: 551-4.
 32. Cozar JM, Romero JM, Aptsiauri N, Vazquez F, Vilchez JR, Tallada M, et al. High incidence of CTLA-4 AA (CT60) polymorphism in renal cell cancer. *Hum Immunol*. 2007; 68: 698-704.
 33. Monne M, Piras G, Palmas A, Arru L, Murineddu M, Latte G, et al. Cytotoxic T-lymphocyte antigen-4 (CTLA-4) gene polymorphism and susceptibility to non-Hodgkin's lymphoma. *American Journal of Hematology*. 2004; 76: 14-8.
 34. Wong YK, Chang KW, Cheng CY, Liu CJ. Association of CTLA-4 gene polymorphism with oral squamous cell carcinoma. *Journal of Oral Pathology & Medicine*. 2006; 35: 51-4.
 35. Haimila K, Turpeinen H, Alakulppi NS, Kyllonen LE, Salmela KT, Partanen J. Association of Genetic Variation in Inducible Costimulator Gene With Outcome of Kidney Transplantation. *Transplantation*. 2009; 87: 393-6.
 36. Magistrelli G, Jeannin P, Herbault N, de Coignac AB, Gauchat JF, Bonnefoy JY, et al. A soluble form of CTLA-4 generated by alternative splicing is expressed by nonstimulated human T cells. *European Journal of Immunology*. 1999; 29: 3596-602.
 37. Shih SC, Yang HW, Chang TY, Hu KC, Chang SC, Lin CL, et al. Investigation of cytotoxic T-lymphocyte-associated protein 4 gene polymorphisms in symptomatic gallstone disease. *Hum Immunol*. 2011; 72: 355-8.
 38. Alegre ML, Noel PJ, Eisfelder BJ, Chuang E, Clark MR, Reiner SL, et al. Regulation of surface and intracellular expression of CTLA4 on mouse T cells. *Journal of Immunology*. 1996; 157: 4762-70.
 39. Karimi MH, Motazedian M, Abedi F, Yaghoobi R, Geramizadeh B, Nikeghbalian S. Association of genetic variation in co-stimulatory molecule genes with outcome of liver transplant in Iranian patients. *Gene*. 2012; 504: 127-32.