

## Research Paper

# Relationship of Genetic Polymorphisms of Aldosterone Synthase Gene *Cytochrome P450 11B2* and *Mineralocorticoid Receptors* with Coronary Artery Disease in Taiwan

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## Abstract

The aldosterone synthase gene, *cytochrome P450 11B2* (*CYP11B2*), and *mineralocorticoid receptor* (*MR*) genes have been reported to be associated with coronary artery disease (CAD). In this study, we investigated the association of single nucleotide polymorphisms (SNPs) of *CYP11B2* (*CYP11B2* T-344C) and *MR* (*MR* C3514G and *MR* C4582A) with CAD in Taiwanese. Six hundred and nine unrelated male and female subjects who received elective coronary angiography were recruited from Chung Shan Medical University Hospital. The enrolled subjects were those who had a positive noninvasive test. *CYP11B2* T-344C, *MR* C3514G and *MR* C4582A were determined by polymerase chain reaction-restriction fragment length polymorphism. We found that women with *CYP11B2* C/C had a higher risk of developing CAD. However, there were no significant differences in the genotype distributions of *MR* C3514G and *MR* C4582A between the women with and without CAD. In multivariate analysis, *CYP11B2* T-344C was most significantly associated with CAD in Taiwanese women. In conclusions, *CYP11B2* C/C was more significantly associated with the development of CAD than diabetes mellitus or hypertension. This implies that *CYP11B2* C/C plays a more important role than some conventional risk factors in the development of CAD in Taiwanese women.

Key words: aldosterone synthase gene, *cytochrome P450 11B2*, mineralocorticoid receptors, single nucleotide polymorphism, coronary artery disease, Taiwan women.

## Introduction

Coronary heart disease is a major cause of mortality and morbidity worldwide affecting millions of people. The causes of coronary heart disease are multifactorial and include conventional and nonconventional factors (1, 2). Male gender, hypertension, smoking, hyperlipidemia, and diabetes mellitus (DM) are conventional risk factors, however, nonconventional risk factors have not yet to be well-defined.

The renin-angiotensin-aldosterone system (RAAS), which affects circulatory homeostasis, regu-

lates the functions of cardiovascular, renal and adrenal glands by regulating blood pressure, fluid and sodium balance (3). RAAS maintains blood pressure through its effect on the kidneys to regulate sodium and water balance, and on peripheral blood vessels to increase systemic vascular resistance (4). Abnormal activity of the RAAS may lead to an array of cardiovascular events such as atherosclerotic coronary artery disease (CAD), plaque rupture and myocardial infarction (3, 5). Local aldosterone synthesis may also

play a pathogenic role (6). Renin cleaves angiotensinogen that is synthesized and secreted by the liver to angiotensin I. Circulating angiotensin I is then hydrolyzed to angiotensin II by angiotensin-converting enzyme that is located primarily in the pulmonary and renal endothelium. Angiotensin II initiates a vasoconstrictor response and stimulates aldosterone synthesis by the adrenal glands (7). Aldosterone has been linked to the development of left ventricular cardiac and systemic vascular remodeling, and left heart failure (8, 9). Aldosterone is also known to play an important role in the regulation of blood pressure, cardiac and perivascular fibrosis, increased left ventricular mass and cardiovascular events (10). It is either causative or a disease modifier that facilitates adaptive cardiovascular remodeling (8, 9). Aldosterone acts via binding to the mineralocorticoid receptor (MR) (11).

Aldosterone secretion is regulated largely by the expression level of the final enzyme required for its biosynthesis, aldosterone synthase, which is encoded by the aldosterone synthase gene, *cytochrome P450 11B2* (*CYP11B2*). Aldosterone, or activation of its receptor, MR, has several extra-renal effects that are largely detrimental in the setting of heart disease (12, 13). Because *CYP11B2* and its receptor are implicated in the development of cardiovascular diseases and the SNPs were associated with heart disease (16), we hypothesized that *CYP11B2* single nucleotide polymorphism (SNP *CYP11B2* T-344C) and MR SNPs (*MR C3514G* and *MR C4582A*) would be associated with CAD. To the best of our knowledge, few studies have investigated the roles of *CYP11B2* T-344C, *MR C4582A* or *MR C3514G* in the development of CAD in Taiwan. The aims of this study were to investigate the correlations of *CYP11B2* T-344C, *MR C4582A* and *MR C3514G* with CAD in Taiwanese.

## Materials and methods

### Subjects

Six hundred and nine unrelated male and female subjects who received elective coronary angiography in Chung Shan Medical University Hospital from April 2007 to March 2009 were recruited. The studied population who received coronary angiography included the subjects who had positive noninvasive test such as the treadmill test, myocardial perfusion scan, or cardiac computed tomography scan. All participants received echocardiographic examinations (Philips Healthcare, SONOS 7500) during their clinic visit. The exclusion criteria included patient refusal, known cerebrovascular attack history, peripheral arterial disease, and incomplete medical chart data. The left ventricular mass (LVM) was calculated using the

formula defined by the American Society of Echocardiography:  $0.8 \times \{1.04 \times [(IVSTD + LVEDD + PWTD)^3 - (LVEDD)^3]\} + 0.6$  g, where IVSTD is interventricular septum thickness in diastole, LVEDD is left ventricular end-diastolic dimension, and PWTD is posterior wall thickness in diastole (15). CAD was defined as more than 50% stenosis over any segment of the coronary artery by angiography, a diagnostic gold standard. The collected data included gender, age, co-morbidities such as hypertension and DM, and echocardiographic measurements including LVM, LVEDD and left ventricular end-systolic diameter (LVESD). The study was approved by the Institutional Review Board of Chung Shan Medical University Hospital (CSMUH No: CS07095), and informed consent was obtained from each participant.

### Blood sample collection and genomic DNA extraction

Venous blood was drawn from each subject into Vacutainer tubes containing EDTA and stored at 4°C. Genomic DNA was extracted using QIAamp DNA blood mini kits (Qiagen, Valencia, CA, USA) according to the manufacturer's instructions. The DNA was dissolved in TE buffer [10 mM Tris (pH 7.8), 1mM EDTA] and then quantitated by measurements at an optical density of 260 nm. The final preparation was stored at -20°C and used as templates for polymerase chain reaction.

### Selection of *CYP11B2* T-344C, *MR C3514G* and *MR C4582A* Polymorphisms

We included the *CYP11B2* T-344C SNP in the promoter region which was found to affect the production of *CYP11B2* in a Chinese population (16). Furthermore, the SNPs *MR C3514G* and *MR C4582A* were selected in this study because the gene polymorphism of the SNP has been found to associate with heart disease (14).

### Polymerase chain reaction-restriction fragment length polymorphism (PCR-RFLP)

The SNPs *CYP11B2* T-344C, *MR C3514G*, and *MR C4582A* were determined by PCR-RFLP assay as previously described (14, 17). The primer sequences and restriction enzyme for analysis of the *CYP11B2* T-344C, *MR C3514G*, and *MR C4582A* gene polymorphisms are described in Table 1. The PCR was performed in a 10 µL volume containing 100 ng DNA template, 1.0 µL of 10 × PCR buffer (Invitrogen, Carlsbad, CA), 0.25 U of Taq DNA polymerase (Invitrogen, Carlsbad, CA), 0.2 mM dNTPs (Promega, Madison, WI), and 200 nM of each primer (MDBioInc, Taipei). The Taq DNA polymerase is a relatively low replication fidelity enzyme. To prevent an error oc-

curing, triple experiments were performed in amplification. The PCR cycling conditions were 5 minutes at 94°C followed by 35 cycles of 1 minute at 94°C, 1 minute at 60°C, and 2 minutes at 72°C, with a final step at 72°C for 20 minutes to allow for complete extension of all PCR fragments. A 10 µL aliquot of PCR product was subjected to digestion at 37°C for 4 hours in a 15 µL reaction containing 5 U of restriction enzyme (New England Biolabs, Beverly, MA) and 1.5 µL buffer (New England Biolabs). Digested products were separated on a 3% agarose gel and then stained with ethidium bromide.

**Table 1.** Primer sequences and PCR-RFLP conditions for amplification of *CYP11B2* and *MR* SNPs.

SNP	Sequences	Product	Enzyme
<i>CYP11B2</i> T-344C	5'-CAGGAGGAGACCCCATGTGAC-3' 5'-CCTCCACCCITGTTACAGCC-3'	T/T: 274 bp, 138 bp, 126 bp C/C: 202 bp, 138 bp, 126 bp, 71 bp	<i>Hae</i> III
<i>MR</i> C3514G	5'-AATCGTCTCTCCACTGCTGTA-3' 5'-CAATGCTGGAATAGCTGCT-3'	C/C: 255 bp G/G: 150 bp, 105 bp	<i>Ban</i> II
<i>MR</i> C4582A	5'-TTGGAAAGCCTGCCTCGTT-3' 5'-TCCTGCCATGATCTGTGCGTT-3'	A/A: 286 bp C/C: 286 bp, 194 bp, 92 bp	<i>Msp</i> A1I

## Statistical analysis

Chi-square and Fisher's exact tests were used to examine the relationships between clinical characteristics and the genotype frequencies of *CYP11B2* T-344C, *MR* C3514G and *MR* C4582A with CAD. The Student *t* test and analysis of variance (ANOVA) with post hoc Scheffe test were used to compare the cardiographic measurements between the subjects with and without CAD as well as among the subjects with different genotypes of the *CYP11B2* SNP. Multivariate analysis of the genotype distribution of *CYP11B2* T-344C and clinical variables for their relationships with CAD was performed using a logistic regression model after controlling for variable parameters. A significant difference was defined as a *P* value of less than 0.05. All statistical analyses were performed using SPSS statistical software (version 11.0; SPSS, Inc., Chicago, IL). Odds ratios (ORs) and the 95% confidence intervals (CIs) were estimated using WinPepi software version 10.0 and SPSS.

## Results

The clinical characteristics of the enrolled individuals are shown in Table 2. Of the 609 subjects, 423 individuals were male and 186 female, and 417 had CAD and 192 did not. There were no significant differences in age, LVM, LVEDD and LVESD between

the two groups. The patients with DM and hypertension had a higher risk of developing CAD [*P*<0.001; OR: 1.96, 95% CI: 1.35-2.85; and *P*=0.007; OR: 2.01, 95% CI: 1.33-3.03, respectively] (Table 2).

**Table 2.** Relationships between clinical variables and coronary artery disease (CAD)

Variables	Negative CAD (N=192)	Positive CAD (N=417)	Odds ratio and 95% confidence interval	<i>P</i> value
<b>Race</b>	Taiwanese	Taiwanese		
<b>Residence</b>	Mid-Taiwan	Mid-Taiwan		
<b>Gender</b>				<0.001 <sup>a</sup>
male	114	309	Reference	
female	78	108	0.51 (0.35-0.75)	
<b>Age (years)</b>	66.9±11.6	65.9±11.5		0.314
<b>Diabetes mellitus</b>				<0.001 <sup>a</sup>
negative	129	213	Reference	
positive	63	204	1.96 (1.35-2.85)	
<b>Hypertension</b>				0.007 <sup>a</sup>
negative	59	86	Reference	
positive	113	331	2.01 (1.33-3.03)	
<b>Left ventricular mass (g)</b>	193.48±35.73	197.61±39.10		0.275
<b>left ventricular end-diastolic diameter (mm)</b>	50.11±5.34	49.87±5.58		0.664
<b>left ventricular end-systolic diameter (mm)</b>	34.95±6.10	35.31±6.39		0.568

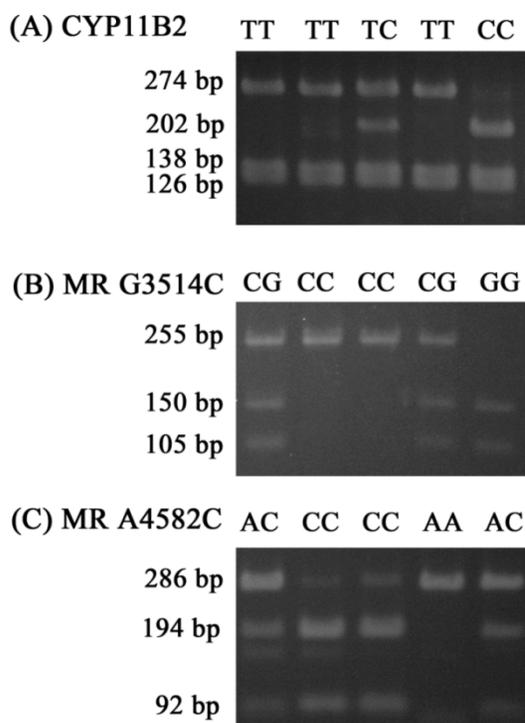
Statistical analysis: Chi-square or independent Student *t* tests

<sup>a</sup>*P*<0.05.

SD: standard deviation.

For the *CYP11B2* gene polymorphism, the wild homozygous alleles (T/T) yielded 274-, 138- and 126-base pair (bp) products, the heterozygous alleles (T/C) yielded 274-, 202-, 138-, 126- and 71-bp products, while the mutant homozygous alleles (C/C) yielded 202-, 138-, 126- and 71-bp products. For *MR* C3514G, the wild homozygous alleles (C/C) yielded a 255-bp product, the heterozygous alleles (C/G) yielded 255-, 150- and 105-bp products, while the mutant homozygous alleles (G/G) yielded 150- and 105-bp products. For *MR* C4582A, the wild homozygous alleles (C/C) yielded 194- and 92-bp products, the heterozygous alleles (C/A) yielded 286-, 194- and 92-bp products, while the mutant homozygous alleles (A/A) yielded a 286-bp product (Fig. 1).

The minor allele frequencies of *CYP11B2* T-344C, *MR* C3514G and *MR* C4582A of the subjects without CAD were all >5% (28.4%, 20.1% and 15.6%, respectively). In these subjects, the genotype frequency of *CYP11B2* (*P*=0.279,  $\chi^2$  value: 4.12) met Hardy-Weinberg equilibrium. The frequencies of *MR* C3514C (*P*>0.05,  $\chi^2$  value: 0.018) and *MR* C4582A (*P*=0.851,  $\chi^2$  value: 0.59) were also in Hardy-Weinberg equilibrium.



**Figure 1.** Polymerase chain reaction-restriction fragment length polymorphisms of *CYP11B2* T-344C, *MR G3514C*, and *MR A4582C* genes. (A) PCR products of *CYP11B2* T-344C gene polymorphisms were subjected to enzymatic digestion by incubation with Hae III, for 4 hours at 37°C and then submitted to electrophoresis in 3% agarose gels. The wild homozygous alleles (T/T) yielded 274-, 138- and 126-base pair (bp) products, the heterozygous alleles (T/C) yielded 274-, 202-, 138-, 126- and 71-bp products, while the mutant homozygous alleles (C/C) yielded 202-, 138-, 126- and 71-bp products. (B) PCR products of the *MR G3514C* gene polymorphism were subjected to enzymatic digestion by incubation with Ban II. The wild homozygous alleles wild (C/C) yielded a 255-bp product, the heterozygous alleles (C/G) yielded 150- and 105-bp products, while the mutant homozygous alleles (G/G) yielded 150- and 105-bp products. (C) PCR products of the *MR A4582C* gene polymorphism were subjected to enzymatic digestion by incubation with MspAII. The wild homozygous alleles (C/C) yielded 194- and 92-bp products, the heterozygous alleles (C/A) yielded 286-, 194- and 92-bp products, while the mutant homozygous alleles (A/A) yielded a 286-bp product.

There were no significant differences in the genotype distributions of *CYP11B2* T-344C, *MR C3514G* and *MR C4582A* SNPs between the subjects with and without CAD (Table 3). When stratified by the gender, these findings remained insignificant in the male subgroup (Table 4). The female subjects with *CYP11B2* C/C had a higher risk of developing CAD, however this risk was not found in the women who had only one mutant allele C (heterozygous T/C) (Table 5). There were no significant differences in the genotype distributions of *MR C3514G* and *MR C4582A* SNPs between the women with and without CAD (Table 5). In addition, we also found that women with DM had a tendency to develop CAD ( $P=0.042$ ; OR: 1.85, 95% CI: 0.98-3.53; Table 5). The women with hypertension had a higher risk of developing CAD ( $P=0.016$ ; OR: 2.44, 95% CI: 1.10-5.48) (Table 6). In multivariate analysis we found that the *CYP11B2* T-344C SNP and hypertension were significantly associated with the development of CAD in the female

subjects ( $P<0.001$  OR:  $\infty$ , 95% CI:  $>1.23-\infty$  and  $P=0.021$ , OR: 2.51, 95% CI: 1.14-5.56, respectively; Table 6).

We next investigated the association of the *CYP11B2* T-344C SNP with cardiographic measurements, and found that the women with *CYP11B2* C/C had a significantly higher LMV compared to those with T/T ( $237.90\pm54.16$  vs.  $189.45\pm38.30$  g,  $P=0.022$ ) and those with T/T or T/C ( $237.90\pm54.16$  vs.  $192.02\pm40.10$  g,  $P=0.005$ ; Table 7). Of the women with CAD, those with *CYP11B2* C/C had a significantly higher LMV compared to those with T/T ( $237.90\pm54.16$  vs.  $188.83\pm41.85$  g,  $P=0.027$ ) and those with T/T or T/C ( $237.90\pm54.16$  vs.  $187.73\pm39.90$  g,  $P=0.005$ ). The women having CAD with the mutant homozygous CC also exhibited a significantly greater LVEDD compared to those with T/T or T/C ( $52.48\pm2.60$  vs.  $47.93\pm4.84$  mm,  $P=0.026$ ). Regardless of the presence of CAD, *CYP11B2* C/C seemed to exacerbate the left ventricle function in the female subjects. However LVM and LVEDD were not associated with the development of CAD in the women (Women with CAD vs. those without CAD: for LVM,  $191.40\pm42.76$  vs.  $192.88\pm40.50$  g,  $P=0.835$ ; for LVEDD,  $48.26\pm4.85$  vs.  $49.16\pm5.10$  mm,  $P=0.285$ ; for LVESD,  $33.81\pm5.34$  vs.  $33.67\pm4.41$  mm,  $P=0.866$ ). This implies that *CYP11B2* C/C but not LMV or LVEDD predispose Taiwanese women to CAD.

**Table 3.** Genotype distributions of single nucleotide polymorphisms of aldosterone synthase gene, *cytochrome P450 11B2* (*CYP11B2*), *CYP11B2* T-344C and *mineralocorticoid receptor* (*MR C3514G* and *MR C4582A*) in subjects with or without coronary artery disease (CAD)

Variables	Negative CAD (N=192)	Positive CAD (N=417)	Odds ratio and 95% confidence interval	P value
<b><i>CYP11B2</i> T-344C</b>				
T/T <sup>a</sup>	93	222	Reference	0.090
T/C	89	159	0.75 (0.52-1.08)	
C/C	10	36	1.51 (0.70-3.55)	
T/T <sup>a</sup>	93	99	Reference	0.271
T/C and C/C	222	195	0.83 (0.58-1.18)	
T/T and T/C <sup>a</sup>	182	381	Reference	0.137
C/C	10	36	1.72 (0.81-3.97)	
<b><i>MR C3514G</i></b>				
C/C <sup>a</sup>	123	267	Reference	0.888
C/G	61	129	0.97 (0.66-1.44)	
G/G	8	21	1.21 (0.50-3.25)	
C/C <sup>a</sup>	123	267	Reference	0.994
C/G and G/G	69	150	1.00 (0.69-1.46)	
C/C and C/G <sup>a</sup>	184	396	Reference	0.640
G/G	8	21	0.98 (0.41-2.62)	
<b><i>MR C4582A</i></b>				
C/C <sup>a</sup>	138	270	Reference	0.116
C/A	48	138	1.47 (0.98-2.22)	
A/A	6	9	0.77 (0.24-2.68)	
C/C <sup>a</sup>	138	270	Reference	0.082
C/A and A/A	54	147	1.39 (0.94-2.06)	
C/C and C/A <sup>a</sup>	186	408	Reference	0.574
A/A	6	9	0.68 (0.21-2.37)	

Statistical analysis: Chi-square or Fisher's exact tests

<sup>a</sup>Used as references for comparison to evaluate the odds ratio of other genotypes.

**Table 4.** Relationships of genotype distribution of single nucleotide polymorphisms of *cytochrome P450 11B2 (CYP11B2 T-344C)* and *mineralocorticoid receptor (MR C3514G and C4582A)* with coronary artery disease (CAD) in Taiwanese men (N=423).

Variables	Negative CAD (N=114)	Positive CAD (N=309)	Odds ratio (OR) and 95% confidence interval	P value <sup>a</sup>
<b>CYP11B2 T-344C</b>				
T/T <sup>b</sup>	50	162	Reference	0.285 <sup>a</sup>
T/C	54	122	0.70 (0.44-1.10)	
C/C	10	25	0.77 (0.35-1.72)	
T/T <sup>b</sup>	50	162	Reference	0.118
T/C and C/C	64	147	0.71 (0.46-1.09)	
<b>MR C3514G</b>				
C/C <sup>b</sup>	77	197	Reference	0.658
C/G	31	98	1.24 (0.76-2.00)	
G/G	6	14	0.91 (0.34-2.46)	
C/C <sup>b</sup>	77	197	Reference	0.469
C/G and G/G	37	112	1.18 (0.75-1.87)	
<b>MR C4582A</b>				
C/C <sup>b</sup>	80	201	Reference	0.456
C/A	30	100	1.33 (0.82-2.15)	
A/A	4	8	0.80 (0.23-2.72)	
C/C <sup>b</sup>	80	201	Reference	0.322
C/A and A/A	34	108	1.26 (0.80-2.01)	

Statistical analysis: Chi-square or Fisher's exact tests

<sup>a</sup>P<0.05.

<sup>b</sup>Used as references for comparison to evaluate the odds ratio of other genotypes.

**Table 5.** Relationships of genotype distribution of single nucleotide polymorphisms of *cytochrome P450 11B2 (CYP11B2 T-344C)* and *mineralocorticoid receptor (MR C3514G and C4582A)* with coronary artery disease (CAD) in Taiwanese women (N=186)

Variables	Negative CAD (N=78)	Positive CAD (N=108)	Odds ratio (OR) and 95% confidence interval	P value <sup>a</sup>
<b>CYP11B2 T-344C</b>				
T/T <sup>b</sup>	43	60	Reference	0.010 <sup>a</sup>
T/C	35	37	0.76 (0.40-1.45)	
C/C	0	11	∞ (1.67-∞)	
T/T <sup>b</sup>	43	60	Reference	0.954
T/C and C/C	35	48	0.98 (0.53-1.84)	
T/T and T/C <sup>b</sup>	78	97	Reference	0.003 <sup>a</sup>
C/C	0	11	∞ (1.93-∞)	
<b>MR C3514G</b>				
C/C <sup>b</sup>	46	70	Reference	0.223
C/G	30	31	0.68 (0.35-1.33)	
G/G	2	7	2.30 (0.41-23.51)	
C/C <sup>b</sup>	46	70	Reference	0.417
C/G and G/G	32	38	0.78 (0.41-1.49)	
C/C and C/C <sup>b</sup>	76	101	Reference	0.308
G/G	2	7	2.63 (0.48-26.56)	
<b>MR C4582A</b>				
C/C <sup>b</sup>	58	69	Reference	0.158
C/A	18	38	1.77 (0.88-3.66)	
A/A	2	1	0.42 (0.01-8.32)	
C/C <sup>b</sup>	58	69	Reference	0.130
C/A and A/A	20	39	1.64 (0.83-3.30)	
C/C and C/A <sup>b</sup>	76	107	Reference	0.573
A/A	2	1	0.36 (0.01-6.97)	

Statistical analysis: Chi-square or Fisher's exact tests

<sup>a</sup>P<0.05.

<sup>b</sup>Used as references for comparison to evaluate the odds ratio of other genotypes.

**Table 6.** Univariate and multivariate analyses of genotype distributions of single nucleotide polymorphisms of *cytochrome P450 11B2 (CYP11B2 T-344C)* and clinical variables for coronary artery disease (CAD) in Taiwanese women

Univariate analysis	Negative CAD (N=78)	Positive CAD (N=108)	OR and 95% CI	P value <sup>a</sup>
<b>CYP11B2 T-344C</b>				
T/T and T/C <sup>b</sup>	78	97	Reference	0.003 <sup>a</sup>
C/C	0	11	∞ (1.93-∞)	
<b>Diabetes mellitus</b>				
negative <sup>b</sup>	50	53	Reference	0.042 <sup>a</sup>
positive	28	55	1.85 (0.98-3.53)	
<b>Hypertension</b>				
negative <sup>b</sup>	22	15	Reference	0.016 <sup>a</sup>
positive	56	93	2.44 (1.10-5.48)	
<b>Multivariate analysis</b>				
<b>CYP11B2 T-344C</b>				
T/T and T/C <sup>b</sup>	78	97	Reference	P value <sup>a</sup>
C/C	0	11	∞ (>1.23-∞)	<0.001 <sup>a</sup>
<b>Diabetes mellitus</b>				
negative <sup>b</sup>	50	53	Reference	0.097
positive	28	55	1.69 (0.91-3.16)	
<b>Hypertension</b>				
negative <sup>b</sup>	22	15	Reference	0.021 <sup>a</sup>
positive	56	93	2.51 (1.14-5.56)	

Statistical analysis: univariate analysis using the chi-square or Fisher's exact tests; multivariate analysis using a logistic regression model after controlling for CYP11B2, diabetes mellitus and hypertension.

<sup>a</sup>P<0.05.

<sup>b</sup>Used as references.

**Table 7.** Relationships of genotype distributions of single nucleotide polymorphisms of *cytochrome P450 11B2 (CYP11B2 T-344C)* with cardiographic measurements in Taiwanese women (N=186).

Variables	LVM (g)	P value <sup>a</sup>	LVEDD (mm)	P value <sup>a</sup>	LVESD (mm)	P value <sup>a</sup>
<b>CYP11B2 T-344C</b>						
T/T	189.45±38.30	0.022 <sup>a</sup>	48.54±5.16	0.170	33.72±5.23	0.403
T/C	190.85±42.92	0.030 <sup>a</sup>	48.39±4.73	0.158	33.50±4.67	0.359
C/C <sup>b</sup>	237.90±54.16		52.48±2.60		36.55±2.79	
T/T and T/C	190.02±40.10	0.005 <sup>a</sup>	48.48±4.97	0.052	33.63±4.99	0.158
C/C <sup>b</sup>	237.90±54.16		52.48±2.60		36.55±2.79	

Statistical analysis: analysis of variance (ANOVA) with post hoc Scheffe test.

LVM: left ventricular mass; LVEDD: left ventricular end-diastolic diameter; LVESD: left ventricular end-systolic diameter; SD: standard deviation.

<sup>a</sup>P<0.05.

<sup>b</sup>Genotype C/C was compared with other genotypes.

## Discussion

This study showed that patients with DM and hypertension had a higher risk of developing CAD. This risk was still present in the female subgroup after stratification by gender. Hypertension and DM, which are conventional risk factors, occurred more frequently in the subjects with CAD. In the Framingham Heart Study, high-normal blood pressure (defined as a systolic blood pressure of 130-139 mmHg, diastolic blood pressure of 85-89 mmHg, or both) increased the risk of cardiovascular disease by 2-fold compared

with healthy individuals (18). Patients with DM have been reported to be 2 to 8 times more likely to experience future cardiovascular events than age- and ethnically-matched individuals without DM (19). However, multivariate analysis in the current study showed that hypertension but not DM was significantly associated with the development of CAD in Taiwanese women.

We conducted this study to define the relationship of a nonconventional risk factor, genetic polymorphism, with CAD in Taiwanese. We found no significant differences in the genotype distributions of *CYP11B2* T-344C, *MR* C3514G and *MR* C4582A SNPs between the subjects with and without CAD. When stratified by gender, the findings remained insignificant in the male subgroup. However, the women with *CYP11B2* C/C had a higher risk of developing CAD, although this risk was not found in the women who had only one mutant allele C. There were no significant differences in the genotype distributions of *MR* C3514G and *MR* C4582A SNPs between the women with and without CAD. A common single nucleotide polymorphism, T to C transition for position -344, occurs within the promoter region of *CYP11B2* (20). In an in vitro study, the C allele was found to bind steroidogenic transcription factor 1 four times more than the T allele (21), and it has also been linked to increased aldosterone production (22, 23). The *CYP11B2* promoter polymorphism has been linked to hypertension (24), and the -344C allele in particular to the risk of acute myocardial infarction (25). In a study of angiotensin II receptor blockers, the CC genotype was found to significantly predict a positive response to antihypertensive treatment (26). However, an association of the -344 genotype with aldosterone levels has been inconsistent, with several studies reporting an association between the -344T allele and higher levels (15, 27). Moreover, a meta-analysis suggested that the -344T>C polymorphism in the *CYP11B2* gene might be associated with susceptibility to CAD in Caucasians and Asians (28). However without stratification by the gender, Mishra et al. reported that *CYP11B2* was not associated with either CAD or left ventricular dysfunction in an Indian population (29).

Even when stratified by gender, the patients with the *MR* C3514G and *MR* C4582A SNPs were still not associated with CAD in our study. This may be due to not specific enough binding of MR with its ligands. MR can bind cortisol and aldosterone with nearly equal affinity (30). Hudson et al. demonstrated the structure of the human MR DNA binding domain in complex with a canonical DNA response element. The overall structure is similar to the glucocorticoid receptor DNA binding domain, however small changes in the mode of DNA binding and lever arm

conformation may explain the differential effects on gene regulation by the mineralocorticoid and glucocorticoid receptors (31). Glucocorticoids activate MR in most tissues at basal levels and glucocorticoid receptors at stress levels (32). Inactivation of cortisol and corticosterone by 11 $\beta$ -hydroxysteroid dehydrogenase allows aldosterone to activate MR within aldosterone target cells and limits the activation of glucocorticoid receptors. Genetic polymorphisms of the *MR* gene could potentially affect both cortisol- and aldosterone-mediated MR effects in the brain and kidneys, respectively (33), which may then complicate the role of MRs in CAD. In addition, Sia et al. revealed no significant differences in the genetic distribution of *MR* between normotensive and hypertensive patients, nor were there differences in the echocardiographic measurements (34).

A report on the Framingham study suggests that variance in aldosterone levels is primarily due to non-genetic factors (35). However, we examined the genetic polymorphism of *CYP11B2*, the gene responsible for aldosterone synthase, in subjects who received coronary catheterization in Taiwan, and found that the C/C allele occurred more frequently in females who had CAD, and that it was associated with higher LVM and LVEDD. In contrast, no C/C alleles were detected in the women who did not have CAD. These results suggest that a genetic variation in aldosterone production may lead to a different prognosis. Bress et al., Takai et al. and Pojoga et al. found that the *CYP11B2* -344C/C genotype was over-represented among individuals with extreme elevation of aldosterone in patients with dilated cardiomyopathy or cardiovascular diseases (36, 37). The association of the *CYP11B2* -344CC genotype with high serum aldosterone levels may explain the reported association between this SNP and greater LVM and decreased event-free survival among African Americans with heart failure (36, 38). In the current study, *CYP11B2* C/C predisposed Taiwanese women to CAD. Regardless of the presence of CAD, *CYP11B2* C/C exacerbated left ventricle function including LVM and LVEDD in the Taiwanese women; however, LVM and LVEDD were not associated with the development of CAD in these women. In multivariate analysis, *CYP11B2* C/C exhibited a more significant association with and a higher risk of developing CAD than DM or hypertension. This implies that the genetic factor *CYP11B2* C/C plays a more important role than some conventional risk factors and functional parameters for the development of CAD in Taiwanese women. Nevertheless, previous studies on the *CYP11B2* T-344C polymorphism have shown a significant (21, 39) or lack of association with hypertension and other cardiovascular parameters (40). Moreover, Jia et al.

suggested that the -344C allele may be associated with a decreased risk of idiopathic hyperaldosteronism (41). Further studies are warranted to elucidate the role of *CYP11B2* C/C in the development of CAD.

One of the limitations of our study is the low sample size. Furthermore, the level of *CYP11B2* gene of CAD patients versus non-CAD control to see how SNP *CYP11B2* T-344C, in particular, that carrying homozygotic CC mutation, affect *CYP11B2* in atherosclerosis is worth for further investigation, which will be included in our future work.

In conclusion, in the present study, we used the candidate gene approach to determine whether the genetic variants of *CYP11B2* T-344C, *MR* C3514G and *MR* C4582A are important effectors in CAD patients. We found no significant differences in the genotype distributions of *CYP11B2* T-344C, *MR* C3514G and *MR* C4582A SNPs between subjects with and without CAD. When stratified by gender in multivariate analysis, *CYP11B2* T-344C exhibited a strong association with the development of CAD in Taiwanese women.

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## Competing Interests

None declared.

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