Establishment and characterization of the reversibly immortalized mouse fetal heart progenitors

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SUPPLEMENTARY DATA

Table S1. List of PCR primers used in the study

Supplementary Table 1. PCR Primers		
	Forward	**Reverse**
cardiac alpha-actin (Actc1)	CCCTGGATTTTGAGAACGAG	GAGTCTCTGGACAGCGGAAG
alpha MyHC	GAGGACCAGGCCAATGAGTA	GCTGGGTGTAGGAGAGCTTG
ANF	GGGGGTAGGATTGACAGGAT	GGATCTTTTGCGATCTGCTC
c-kit	GGGCTAGCCAGAGACATCAG	AGGAGAAGAGCTCCCAGAGG
cTnT	CTGAGACAGAGGAGGCCAAC	ACCAAGTTGGGCATGAAGAG
Cx43	CCTCAACATGGCATTTCCTT	TCCACACCTAGAGGGTCAGG
GAPDH	GGCTGCCCAGAACATCAT	CGGACACATTGGGGGTAG
GATA4	TCTCCCAGGAACATCAAACC	GTGTGAAGGGGTGAAAAGGA
Hand2	CTACTTCCACGGCTGGCTTA	CCATAATGGGAGTGGTCCAG
ISL-1	AAGGACAAGAAACGCAGCAT	CCATCATGTCTCTCCGGACT
Nanog	AAGTACCTCAGCCTCCAGCA	GTGCTGAGCCCTTCTGAATC
Nkx2.5	GAGCCTGGTAGGGAAAGAGC	GGTGGGTGTGAAATCTGAGG
Oct3/4	CTGGGCGTTCTCTTTGGA	GGCTTCCTCCACCCACTT
Sca-1	CCTGGAGCCCTCTAGTGATG	GAGCAGCAATCCACAACAAA
Sox2	ACAGCTACGCGCACATGA	TGCATCGGTTGCATCTGT
Tbx5	CGCTGTGACTTCGTACCAGA	ACTTTGCATCCGAGACATCC

Figure S1. Characterization of iCP15 clones. Total RNA was isolated from 20 chosen iCP15 clones and subjected to reverse transcriptase reactions and semi-quantitative RT-PCR analysis of marker expression, including the common stem cell markers Oct3/4, Nanog, and Sox2 (**A**), the early cardiomyogenic progenitor markers, ISL-1, Sca1 and c-kit (**B**), the immediate early cardiomyogenic progenitor markers Nkx2.5, GATA4, Tbx5, and Hand2 (**C**), and the late cardiomyogenic markers, cardiac α-actin (Actc1), α-myosin heavy chain (α-MyHC), cardiac troponin T (cTnT) and Atrial natriuretic factor (ANF) (**D**). All samples were normalized for their GAPDH expression. All RT-PCR reactions were repeated at least in three independent experiments. Representative results are shown. P19 is a mouse embryonal carcinoma line and was used as a control line.

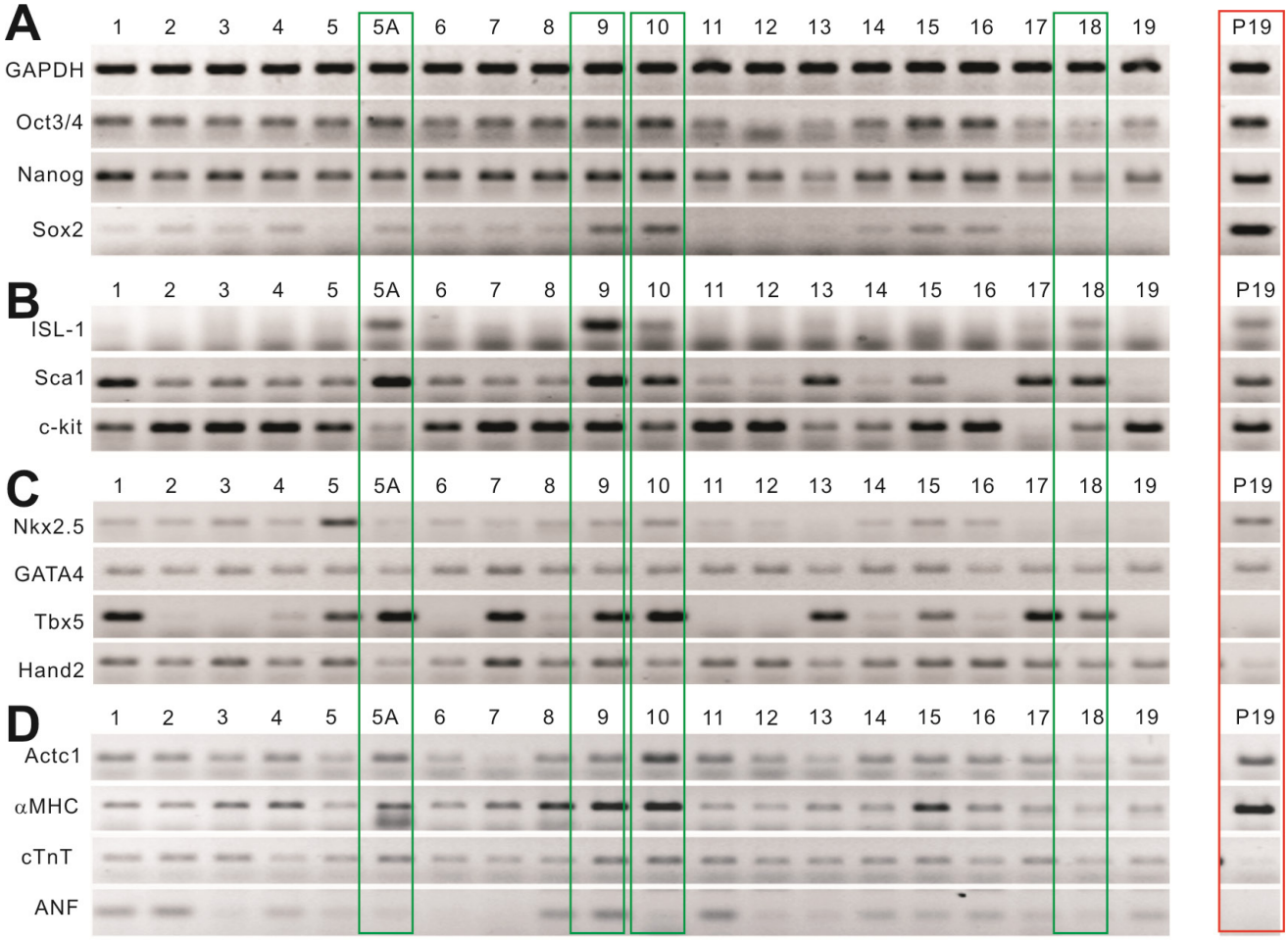

Figure S2. Immunofluorescence staining of cardiomyogenic progenitor markers for the iCP15 clones. The selected iCP15 clones were seeded in subconfluence, fixed and stained with ISL-1 (**A**) or c-kit antibody (**B**), followed by staining with DyLight 488 or DyLight 594-labeled secondary antibody. Isotope IgG was used as staining control.